

Methodology article

Open Access

# A minimally invasive method of piscine tissue collection and an analysis of long-term field-storage conditions for samples

James J Campanella\*<sup>1</sup> and John V Smalley<sup>2</sup>

Address: <sup>1</sup>Montclair State University, Department of Biology and Molecular Biology, 1 Normal Avenue, Upper Montclair, New Jersey 07043, USA and <sup>2</sup>Bergen Community College, Department of Science and Technology, 400 Paramus Road, Paramus, New Jersey 07652, USA

Email: James J Campanella\* - james.campanella@montclair.edu; John V Smalley - jsmalley@bergen.edu

\* Corresponding author

Published: 30 May 2006

Received: 23 March 2006

BMC Genetics 2006, 7:32 doi:10.1186/1471-2156-7-32

Accepted: 30 May 2006

This article is available from: <http://www.biomedcentral.com/1471-2156/7/32>

© 2006 Campanella and Smalley; licensee BioMed Central Ltd.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

## Abstract

**Background:** The acquisition of high-quality DNA for use in phylogenetic and molecular population genetic studies is a primary concern for evolutionary and genetic researchers. Many non-destructive DNA sampling methods have been developed and are used with a variety of taxa in applications ranging from genetic stock assessment to molecular forensics.

**Results:** The authors have developed a field sampling method for obtaining high-quality DNA from sunfish (*Lepomis*) and other freshwater fish that employs a variation on the buccal swab method and results in the collection of DNA suitable for PCR amplification and polymorphism analysis. Additionally, since the circumstances of storage are always a concern for field biologists, the authors have tested the potential storage conditions of swabbed samples and whether those conditions affect DNA extraction and PCR amplification. It was found that samples stored at room temperature in the dark for over 200 days could still yield DNA suitable for PCR amplification and polymorphism detection.

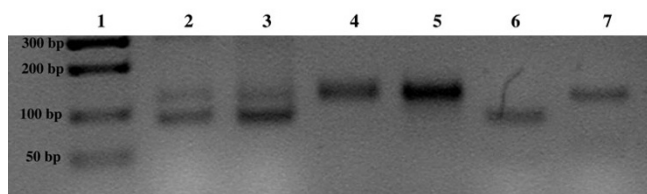
**Conclusion:** These findings suggest that valuable molecular genetic data may be obtained from tissues that have not been treated or stored under optimal field conditions. Furthermore, it is clear that the lack of adequately low temperatures during transport and long term storage should not be a barrier to anyone wishing to engage in field-based molecular genetic research.

## Background

The acquisition of high quality DNA for use in phylogeny and population genetic studies is a prime concern for researchers. While such DNA samples are easily obtained, it often requires the sacrifice of the animals being studied. Such lethal sampling has the potential to seriously impact the genetic makeup of populations under investigation, altering the composition of the future population.

A number of non-destructive nucleic acid sampling methods have been developed with a variety of taxa in applica-

tions ranging from genetic stock assessment to molecular forensics. DNA samples suitable for PCR amplification and analysis of polymorphisms in honey bees (*Apis mellifera*) have been obtained from wing clips [1]. Toe clips have been used to obtain samples of DNA for genetic studies of the Great Plains toad, *Bufo cognatus* [2]. In piscine species, tissue sources of DNA for non-lethal sampling include: fin clips, scales, barbels, muscle, blood, and sperm [3,4]. The molecular phylogeny of the family *Chinichillidae* has been investigated using DNA from hair, blood, feces, and ear tissue [5]. DNA suitable for micros-



**Figure 1**

PCR products of bluegill DNA (Lake Wapalanne), employing Lma20 primers to amplify microsatellite regions. Lane 1: molecular weight markers (Hi-Lo Marker, Minnesota Molecular), size indicated in basepairs. Lanes 2–7: bluegill Lma20 microsatellite polymorphisms. Buccal tissues employed for this experiment were stored overnight in 100% ethanol before DNA extraction after 24 hours. Heterozygotes and homozygotes for the Lma20 marker are clearly delineated in the individual fish. 2% agarose gel stained with ethidium bromide. The image was inverted to a negative by Scion computer software (Scion, Inc., Frederick, Maryland).

atellite analysis and genotyping has even been obtained from Amur tiger (*Panthera tigris altaica*) and chimpanzee (*Pan troglodytes*) feces [6,7], and from long-dead sperm whales (*Physeter macrocephalus*) using teeth and scrimshaw [8].

A standard method of collecting DNA from humans involves the minimally invasive process of buccal swabbing to separate away epithelial cells from which the DNA is then extracted [9,10]. Among the advantages of this method are its swiftness and ease of application [11]. These characteristics make buccal swabbing adaptable to a wide variety of situations and particularly amenable to large sample sizes.

In addition to the problem of collecting tissue samples non-destructively, the storage of said samples in the field for subsequent DNA extraction presents a difficulty for the field biologist involved in molecular analysis. Since elaborate DNA extraction methods that would ensure sample stability usually can not be employed in the field, alternatives must be found. Low temperature tissue storage is an obvious, although potentially impractical, solution. Tissues fixed in 50% ethanol have yielded DNA suitable for restriction endonuclease digestion and hybridization with oligonucleotide probes after storage for 6 years [12]. Storage of tissues in a Lysis/Storage/Transportation (LST) buffer at room temperature for up to 8 weeks has allowed extraction of DNA of sufficient quality for restriction endonuclease digestion as well as PCR amplification [13].

The authors have developed a field sampling method for obtaining high quality DNA from bluegill sunfish (*Lep-*

*omis macrochirus*). This method employs a variation on the buccal swab method and results in the collection of DNA suitable for PCR amplification and polymorphic analysis. Additionally, we address the question of how robust our method would be for a field scientist who did not have the proper storage facilities. If a field scientist did not have proper refrigeration, would tissues sampled by buccal swabbing be stable and, if so, for how long?

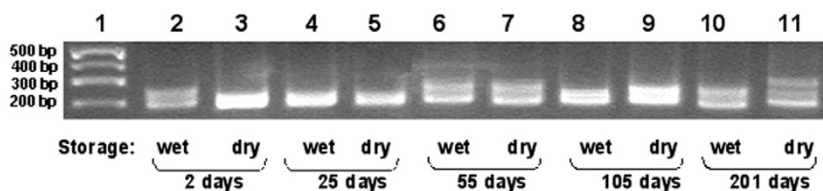
## Results and discussion

The quality of the isolated bluegill DNA is high enough to allow PCR amplification of simple sequence repeat polymorphisms without further purification. There is some background visible along with the polymorphic DNA bands, but the bands themselves are clearly visible for each individual fish (Figure 1). Microsatellite regions have been successfully amplified at several bluegill loci (Lma20, Lma21, Lma87, Lma102, Lma120, and Lma124), although data from only the Lma20 locus is presented here (Figure 1). Moreover, we have amplified larger polymorphic microsatellite sequences of 300 basepairs using the marker Lma120 (data not shown). High-resolution agarose gel electrophoretic analysis using comparison to known concentrations of HiLo Marker (Minnesota Molecular, Madison, Minnesota) was employed to determine the size ranges of unamplified genomic DNA. We found molecular weights ranged from 1000 to 7000 basepairs.

After over 200 days of storage, either preserved in ethanol or in a dried state, the *Lepomis* tissue yielded DNA that amplifies microsatellite sequences quite clearly and strongly (Fig. 2). Additionally, the polymorphisms among the individual bluegills sampled can be identified, suggesting that population studies may be performed even on samples that are months old and stored at room temperature. We conclude that tissue samples taken in this manner are quite stable over long periods either when vacuum dried or left in ethanol.

Note that the PCR products being amplified and examined (Fig. 2) are not from the same individuals, so many of the differences that we are seeing in amplified products arise from polymorphisms between the individuals.

There does appear to be an aberrant product at 201 days between the "wet" and the "dry" preserved samples (Fig. 2, lanes 10 and 11) where an extra band has appeared. There may have been some minor degradation in the dried tissue sample that led to an additional PCR product being generated. It seems more likely that we may be observing two microsatellite (Lma20) alleles, and a heteroduplex band composed of two different size alleles. "Triplet" bands are often produced in PCR, where two basepair repeats are found in the microsatellite sequence,



**Figure 2**

PCR amplification of DNA polymorphisms from aged sunfish tissues, employing Lma20 primers to amplify microsatellite regions. Lane 1: molecular weight markers (Hi-Lo Marker, Minnesota Molecular), size indicated in basepairs. Lanes 2, 4, 6, 8, and 10: PCR amplification of sunfish DNA from tissues that were stored in 100% ethanol. Lanes 3, 5, 7, 9, and 11: PCR amplification of sunfish DNA from tissues that were stored in a dried state. The period of aging is indicated for each set of samples. 2% agarose gel stained with ethidium bromide.

such as in Lma20 (personal communication, Dr. Kirsten Monsen).

The mean concentration of DNA extracted using each method of preservation was comparable: the "wet" preserved DNA was  $50.6 \pm 7.6$  ng/uL, while the "dry" preserved was  $46.4 \pm 5.1$  ng/uL. The range in "dry" concentrations was 29–70 ng/uL. The range of "wet" concentration was 32–69 ng/uL.

It should be noted that although we employed 100% ethanol successfully in our own tissue stability studies, it seems likely that concentrations as low as 50% will work as well [12]. Many researchers may want to consider lowering the ethanol concentration for reasons of economy.

Our findings suggest that valuable molecular genetic data may be obtained not only from piscine tissues that have been stored under optimal conditions, but also from those that have not been stored optimally. Furthermore, it is clear that the lack of adequately low temperatures during transport and long term storage should not be a barrier to anyone wishing to engage in field-based molecular genetic research. Although we have not endeavoured to determine whether tissues gathered in this manner from other aquatic species may have the same longevity of storage, it is likely that tissues obtained from other fish using the described field extraction method will yield robust samples.

## Methods

### Collection and treatment of samples

Bluegill sunfish (*Lepomis macrochirus*) were caught in situ using standard angling equipment and bait of mealworms. The sunfish originated from Lake Wapalanne in northwestern New Jersey. Once caught, the fish were held temporarily (20–90 min) in buckets of lake water.

When ~20 fish were captured, buccal smears were taken from each fish by sterilely swabbing their mouths using the wooden dowel end of sterile cotton-tipped applicators (Moore Medical Corp, New Britain, Connecticut). Cheek cells from the applicator ends were fixed and preserved on site by re-suspension into 100 uL of 100% denatured ethanol (Fisher Scientific, Hanover Park, Illinois) in 1.5 mL microfuge tubes. After taking buccal smears, fish were returned to their lake habitat.

For the study testing the practicality of using buccal swabbing for the tissue collection process, the fixed tissue samples were stored at 4 °C for 24–96 hours before extraction.

For the tissue stability study, half the samples were left in ethanol, sealed with parafilm and stored at a constant 26 °C (Model E-30B constant temperature chamber, Percival Scientific) in the dark. The remainder of the samples were dried for 20 min on a Savant Speedvac vacuum dryer (GMI, Inc., Albertville, Minnesota) set at the lowest drying temperature. These dried samples were then sealed with parafilm and stored with the ethanol preserved samples in the dark at a constant 26 °C. DNA was extracted from the tissues (both ethanol preserved and vacuum dried) after 2, 5, 25, 55, 105, and 201 days.

### DNA extraction and PCR amplification conditions

All ethanol preserved samples were dried for 20 min on a Savant Speedvac vacuum dryer set at the lowest drying temperature. Both "wet" preserved and "dry" preserved tissues were then resuspended in 50 uL of TE and RNase (10 mM Tris-Cl, 1 mM EDTA, pH 8.0, 1 unit RNase per 50 uL aliquot). Tissues were lysed by 5 min incubation at 95 °C, cooled on ice for an additional 5 min incubation, and centrifuged briefly to collect condensation.

The polymerase chain reaction was performed employing *Lepomis* amplification primers for detecting microsatellite polymorphisms [14]. The amplification conditions prin-

cipally followed the directions of Vander Zwan et al [15]. Microsatellites were amplified in 20 µL reactions containing: 29–70 ng fish DNA, 10% ThermoPol buffer (New England Biolabs Inc., Beverly, Massachusetts), 5 pmoles of each primer, 200 µM dNTPs (New England Biolabs Inc., Beverly, Massachusetts), and 1.0 unit Taq polymerase. The primers used for the test were for the Lma20 locus (Lma20F: 5'GGCACTAATCTAATTGTAGCC 3', Lma20R: 5'TTGTGTGTCTGCATTGGAATC 3') [14]. DNA concentrations were determined employing a Nanodrop Spectrophotometer Model ND-1000 (Nanodrop Technologies Inc., Wilmington, Delaware).

All amplification was performed in a Mastercycler Gradient Thermocycler (Eppendorf Inc., Germany). The PCR products were subjected to electrophoresis on a 2% agarose gel in 1X sodium borate buffer [16]. The products on agarose gels were stained with ethidium bromide and imaged using an Ultralum gel documentation system (Ultralum, Inc., Claremont, California) and Scion computer software (Scion, Inc., Frederick, Maryland).

### Authors' contributions

Both JC and JS collected tissue samples, contributed to the experimentation process, and wrote the submitted manuscript.

### Acknowledgements

Foremost, the authors wish to thank the New Jersey School of Conservation for the use of their facilities. They also wish to thank Lisa Campanella for her editorial help with this manuscript. This work was supported by a Sokol grant for undergraduate research.

### References

- Châline N, Ratnieks FLW, Raine NE, Badcock NS, Burke T: **Non-lethal sampling of the honey bee, *Apis mellifera*, DNA using wing tips.** *Apidologie* 2004, **35**:311-318.
- Gonzalez Z, Ray DA, Mcaliley LR, Gray MJ, Perchellet C, Smith LM, Densmore LD: **Five polymorphic microsatellite markers for the Great Plains toad, *Bufo cognatus*.** *Molecular Ecology Notes* 2004, **4**:9-10.
- Cummings SA, Thorgaard GH: **Extraction of DNA from fish blood and sperm.** *Biotechniques* 1994, **17**(3):426-430.
- Wirgin II, Waldman JR: **What DNA can do for you.** *Fisheries* 1994, **19**(7):16-27.
- Spotorno AE, Valladares JP, Marin JC, Palma RE, Zuleta CR: **Molecular divergence and phylogenetic relationships of chinchillids (Rodentia: Chinchillidae).** *Journal of Mammalogy* 2004, **85**(3):384-388.
- Russello MA, Gladyshev E, Miquelle D, Caccone A: **Potential genetic consequences of a recent bottleneck in the Amur tiger of the Russian far east.** *Conservation Genetics* 2004, **5**(5):707-713.
- Vigilant L: **Technical challenges in the microsatellite genotyping of a wild chimpanzee population using feces.** *Evolutionary Anthropology Suppl* 2002, **1**:162-165.
- Pichler FB, Dalebout ML, Baker CS: **Non-destructive DNA extraction from sperm whale teeth and scrimshaw.** *Molecular Ecology Notes* 2001, **1**:106-109.
- Tobal K, Layton DM, Mufti GJ: **Non-invasive isolation of constitutional DNA for genetic analysis.** *Lancet* 1989, **2**:1281-1282.
- Walker AH, Najarian D, White DL, Jaffe JF, Kanetsky PA, Rebbeck TR: **Collection of genomic DNA by buccal swabs for polymerase chain reaction-based biomarker assays.** *Environmental Health Perspectives* 1999, **107**:517-520.
- London SJ, Xia J, Lehman TA, Yang JH, Granada E, Chunhong L, Dubeau L, Li T, David-Beabes GL, Li Y: **Collection of buccal cell DNA in seventh-grade children using water and a toothbrush.** *Cancer Epidemiology Biomarkers & Prevention* 2001, **10**:1227-1230.
- Smith LJ, Braylan RC, Nutkis JE, Edmundson KB, Downing JR, Wakefield EK: **Extraction of cellular DNA from human cells and tissues fixed in ethanol.** *Analytical Biochemistry* 1987, **160**:135-138.
- Muralidharan K, Wemmer C: **Transporting and storing field-collected specimens for DNA without refrigeration for subsequent DNA extraction and analysis.** *Biotechniques* 1994, **17**(3):420-422.
- Neff BD: **Genetic paternity analysis and breeding success in bluegill sunfish (*Lepomis macrochirus*).** *The Journal of Heredity* 2001, **92**(2):111-119.
- Vander Zwan C, Brodie S, Campanella JJ: **The intraspecific phylogenetics of *Arabidopsis thaliana* in world-wide populations.** *Systematic Botany* 2000, **25**(1):47-59.
- Brody JR, Kern SE: **Sodium boric acid: a Tris-free, cooler conductive medium for DNA electrophoresis.** *Biotechniques* 2004, **36**:214-216.

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:  
http://www.biomedcentral.com/info/publishing\_adv.asp

