

RESEARCH ARTICLE

Open Access

Genome-wide association study for intramuscular fat deposition and composition in Nelore cattle

Aline SM Cesar¹, Luciana CA Regitano², Gerson B Mourão¹, Rymer R Tullio², Dante PD Lanna¹, Renata T Nassu², Maurício A Mudado², Priscila SN Oliveira³, Michele L do Nascimento¹, Amália S Chaves¹, Maurício M Alencar², Tad S Sonstegard⁴, Dorian J Garrick⁵, James M Reecy⁵ and Luiz L Coutinho^{1*}

Abstract

Background: Meat from *Bos taurus* and *Bos indicus* breeds are an important source of nutrients for humans and intramuscular fat (IMF) influences its flavor, nutritional value and impacts human health. Human consumption of fat that contains high levels of monounsaturated fatty acids (MUFA) can reduce the concentration of undesirable cholesterol (LDL) in circulating blood. Different feeding practices and genetic variation within and between breeds influences the amount of IMF and fatty acid (FA) composition in meat. However, it is difficult and costly to determine fatty acid composition, which has precluded beef cattle breeding programs from selecting for a healthier fatty acid profile. In this study, we employed a high-density single nucleotide polymorphism (SNP) chip to genotype 386 Nelore steers, a *Bos indicus* breed and, a Bayesian approach to identify genomic regions and putative candidate genes that could be involved with deposition and composition of IMF.

Results: Twenty-three genomic regions (1-Mb SNP windows) associated with IMF deposition and FA composition that each explain $\geq 1\%$ of the genetic variance were identified on chromosomes 2, 3, 6, 7, 8, 9, 10, 11, 12, 17, 26 and 27. Many of these regions were not previously detected in other breeds. The genes present in these regions were identified and some can help explain the genetic basis of deposition and composition of fat in cattle.

Conclusions: The genomic regions and genes identified contribute to a better understanding of the genetic control of fatty acid deposition and can lead to DNA-based selection strategies to improve meat quality for human consumption.

Keywords: Fatty acid, GWAS, *Bos indicus*, Beef, Positional candidate gene

Background

Many consumers associate consumption of fat from beef with coronary heart disease, diabetes and obesity, due to the presence of cholesterol, high concentration of saturated fatty acids (SFA), and low concentration of polyunsaturated fatty acids (PUFA). However, consumption of fatty acids is necessary for human nutrition [1]. Beef has high nutritional value from children to seniors, is a rich source of protein (essential amino acids), iron, zinc, B vitamins and essential polyunsaturated fatty acids such as linoleic and linolenic acid [2]. Beef fat also has a high concentration of monounsaturated fatty acids (MUFA),

whose melting point is low and can reduce the concentration of bad cholesterol (LDL) in blood circulation [3].

The amount of fatty acid and its composition in beef varies by breed, nutrition, sex, age and carcass finishing level [4]. The difficulties associated with determining intramuscular fat (IMF) deposition and composition as well as the limited knowledge on the genetic mechanisms that control these traits has limited genetic progress in the production of healthier beef.

The development of high-density bovine genotyping [5] and their use in genome-wide association studies (GWAS) have allowed identification of genomic regions associated with phenotypes of interest. The technique of GWAS exploits differences in allele frequencies of thousands of polymorphic markers available in unrelated individuals who possess different phenotypes (for example,

* Correspondence: llcoutho@usp.br

¹Department of Animal Science, University of São Paulo, Piracicaba SP 13418-900, Brazil

Full list of author information is available at the end of the article

deposition and composition of intramuscular fat), and leads to the identification of markers associated with a given phenotype [6]. Bayesian approaches have been applied to GWAS to detect significant quantitative trait loci (QTL) for traits of economic importance. One such approach uses multiple regression (evaluating marker effects simultaneously), treating marker effects as random to reduce overestimation bias of significant QTL effects, generating the actual posterior distribution of QTL effects given the data which can provide richer inference than can be obtained by simply constructing p-values as well as providing an alternative to the use of p-values to avoid false positives [7-9].

Brazilian beef is exported and consumed in more than 100 countries [10]. Purebred and crossbred Nellore cattle, which are of *Bos indicus* descent, are the predominant source of beef in Brazil. Previous research has documented that muscle and fat tissues from *Bos indicus* cattle develop in a different manner than in *Bos taurus* breeds [11-14]. However, studies documenting the genetics of fatty acid deposition and composition in *Bos indicus* breeds are limited.

In this study we performed a GWAS using high-density single nucleotide polymorphism (SNP) chips (770 k) and Bayesian methods (Bayes B) to identify genomic regions associated with fat deposition and fatty acid composition (FA) in Nellore beef.

Results and discussion

Intramuscular fat deposition and composition

Modern consumers are concerned with their overall health and often desire to reduce their caloric intake. This has increased the demand for lean meat production with a healthier fatty acid composition, which would comprise a lower proportion of SFA and greater proportion of MUFA. The amount of fat deposition in meat represented as IMF (mean = 2.77%) reported in this feedlot-finished study was greater than for pasture-finished counterparts, as expected, but lower than normally observed in Continental and English breeds [15-17]. Despite of that, IMF observed in this work was within limits of reasonable amount of fat to assure acceptable quality levels for consumers according to Nuernberg et al. [18].

The fatty acid composition observed for the most abundant FAs were: C14:0 at 3.54%, C16:0 at 26.69%, C18:0 at 14.98%, C16:1 cis-9 at 3.31%, C18:0 at 14.98%, C18:1 cis-9 at 37.46%, C18:2 cis-9 cis 12 at 1.60%, SFA at 47.23%, MUFA at 48.34%, and PUFA at 2.87% (Table 1). The proportion of MUFA was higher than SFA, and oleic acid (C18:1 cis-9) was the most abundant single fatty acid (37.46%). The FA composition presented in this work is similar to those reported in the literature for Nellore or other *Bos indicus* breeds [19-21]. This population also presented, in relation to *Bos taurus*

breeds, average composition of fatty acids which is in agreement with reports from the USDA [22]. However, in this study a lower quantity of PUFA was observed in general, but not for C18:2 cis-9 trans-11, consequently a lower ratio of PUFA/SFA (6.08%). Similar MUFA and PUFA results have been reported in previous studies that utilized *Bos indicus* steers [20,23].

The omega 6: omega 3 (n-6:n-3) ratio was high (4.84), which has a beneficial effect on patients with chronic diseases. A ratio of 4:1 has been associated with a 70% decrease in total mortality in the secondary prevention of cardiovascular disease [24]. Omega 6 and 3 fatty acids are essential cell membrane and tissue constituents that are important for many biological functions. These fatty acids are also essential for synthesis of prostaglandins, thromboxane, leukotriene, hydroxyl fatty acids, and lipoxins that are involved with inflammatory response. Humans and other mammals can convert n-6 to n-3 using desaturation enzymes but this conversion is slow and there is competition between n-6 and n-3 fatty acids for the desaturation enzymes [25].

Heritability

Descriptive statistics and heritabilities estimated using a genomic relationship **G** matrix are in Table 1. Heritabilities estimated in this study varied from low (<0.10 for C12:0, C16:0, C18:1 cis-11, C18:1 cis-12, C18:2 cis-9 trans-11, C20:1, C20:3 n-6, C20:4 n-6, C20:5 n-3 and AI, respectively) to moderate (up to 0.29 for IMF, C14:0, C14:1 cis-9, C16:1 cis-9, C17:0, C17:1, C18:0, C18:1 cis-9, C18:1 trans-6, 7, 8, C18:1 trans-10, 11, 12, C18:2 cis 9 cis-12 n-6, C18:2 trans-11 cis-15, C18:3 n-6, C18:3 n-3, C22:5 n-3, C22:6 n-3, SFA, MUFA, PUFA, Sn-3, Sn-6 and n-6:n-3). For C15:0, C18:1 cis-13, C18:1 cis-15, C18:1 trans-16, C20:2 the heritability estimates were zero. In Angus [26] and Japanese Black cattle [27] estimates of heritability for IMF fat deposition and composition traits were higher than in this study. The lower values of heritability reported for this population could be explained by the reduced sample size [28] or lower amount of genetic variation in the population [29].

The estimates of genomic heritability from Bayes B approach (Table 2) were also low to moderate (from 0.09 to 0.46) and generally similar to values obtained in ASReml using a genomic relationship matrix.

Genome wide association studies and genomic regions identified

GWAS and genomic regions identified were reported for those traits that showed genomic heritability ≥ 0.10 : IMF, 33 different FA and ratios, and one FA index. The 1-Mb SNP window regions that explained more than 1% of the genetic variance were used to search for putative candidate genes (PCG) and are presented on Table 3. Therefore,

Table 1 Descriptive statistics, variance components and heritability by GBLUP for IMF deposition and composition in Nellore

Trait	Terminology ¹	N	Mean ± SE ²	Genetic variance	Residual variance	Total variance	h ² ± SE
IMF (%)	Intramuscular fat	382	2.77 ± 0.05	0.196	0.490	0.686	0.29 ± 0.16
C12:0 (mg/mg)	Lauric acid	374	0.07 ± 0.001	0.000039	0.00072	0.000761	0.05 ± 0.09
C14:0	Myristic acid	378	3.54 ± 0.03	0.0530	0.250	0.303	0.17 ± 0.11
C14:1 cis-9	Myristoleic acid	378	0.96 ± 0.01	0.0076	0.041	0.0486	0.16 ± 0.11
C15:0	Pentadecylic acid	378	0.80 ± 0.02	0.0	0.052	0.052	0 ± 0.06
C16:0	Palmitic acid	378	26.69 ± 0.15	0.6070	7.068	7.675	0.08 ± 0.10
C16:1 cis-9	Palmitoleic acid	378	3.31 ± 0.04	0.0640	0.354	0.418	0.15 ± 0.10
C17:0	Margaric acid	378	1.07 ± 0.009	0.0061	0.019	0.0251	0.24 ± 0.15
C17:1	Heptadecenoic acid	378	0.58 ± 0.007	0.0024	0.009	0.0114	0.20 ± 0.12
C18:0	Stearic acid	378	14.98 ± 0.14	1.3380	5.348	6.686	0.20 ± 0.12
C18:1 cis-9	Oleic acid	378	37.46 ± 0.22	2.0720	10.826	12.898	0.16 ± 0.11
C18:1 cis-11	Cis-Vaccenic acid	378	2.98 ± 0.05	0.0850	0.357	0.442	0.02 ± 0.09
C18:1 cis-12	Cis-12 Octadecenoic	377	0.91 ± 0.02	0.0030	0.034	0.037	0.09 ± 0.10
C18:1, cis-13	Cis-13 Octadecenoic	377	0.58 ± 0.008	0.0	0.021	0.021	0 ± 0.06
C18:1 cis-15	Cis-15 Octadecenoic	377	0.06 ± 0.002	0.0	0.0005	0.0005	0 ± 0.06
C18:1 trans-6, 7, 8	Trans-6,7,8 Octadecenoic	378	0.18 ± 0.004	0.0007	0.0055	0.0062	0.11 ± 0.09
C18:1 trans-10, 11, 12	Trans-10,11,12 Octadecenoic	378	1.07 ± 0.02	0.0236	0.0926	0.1162	0.20 ± 0.12
C18:1 trans-16	Trans-16 Octadecenoic	377	0.14 ± 0.003	0.0	0.0023	0.0023	0 ± 0.09
C18:2 cis-9 cis-12 n-6	Linoleic acid	377	1.60 ± 0.03	0.0340	0.239	0.273	0.12 ± 0.10
C18:2 cis-9 trans-11	Vaccenic acid	377	0.21 ± 0.003	0.0001	0.0026	0.0027	0.04 ± 0.09
C18:2 trans-11 cis-15	Octadecenoic acid	374	0.07 ± 0.001	0.00007	0.0004	0.00047	0.13 ± 0.11
C18:3 n-6	α-Linolenic acid	377	0.06 ± 0.001	0.00008	0.0002	0.00028	0.24 ± 0.13
C18:3 n-3	γ-Linolenic acid	376	0.16 ± 0.005	0.00026	0.0017	0.00196	0.13 ± 0.11
C20:1	Eicosanoic acid	374	0.11 ± 0.002	0.00013	0.0014	0.00153	0.09 ± 0.10
C20:2	Eicosadienoic acid	306	0.01 ± 0.0003	0.0	0.00004	0.00004	0 ± 0.08
C20:3 n-6	Eicosatrienoic acid	373	0.11 ± 0.003	0.00018	0.00235	0.00253	0.07 ± 0.09
C20:4 n-6	Arachidonic acid	377	0.36 ± 0.008	0.00221	0.0235	0.0251	0.09 ± 0.10
C20:5 n-3 (EPA)	Eicosapentaenoic acid	376	0.08 ± 0.002	0.00005	0.00135	0.00139	0.04 ± 0.09
C22:5 n-3 (DPA)	Docosapentaenoic acid	377	0.17 ± 0.003	0.00047	0.00387	0.00434	0.11 ± 0.10
C22:6 n-3 (DHA)	Docosahexaenoic acid	364	0.03 ± 0.001	0.00003	0.00018	0.00021	0.13 ± 0.10
SFA ³	Sum of saturated FA	377	47.23 ± 0.23	1.960	15.00	16.960	0.11 ± 0.09
MUFA ³	Sum of monounsaturated FA	377	48.34 ± 0.23	2.367	14.963	17.33	0.14 ± 0.10
PUFA ³	Sum of polyunsaturated FA	377	2.87 ± 0.04	0.097	0.547	0.644	0.15 ± 0.10
n-3 ⁴	Sum of omega-3	377	0.44 ± 0.09	0.00034	0.0016	0.00194	0.17 ± 0.11
n-6 ⁴	Sum of omega-6	377	2.13 ± 0.04	0.00055	0.0031	0.00365	0.15 ± 0.11
PUFA:SFA	Ratio of PUFA to SFA	377	6.08 ± 0.003	0.0623	0.41	0.4723	0.13 ± 0.10
n-6:n-3	Ratio of n-6 to n-3	377	4.84 ± 0.11	0.00395	0.0231	0.02705	0.14 ± 0.10
AI ⁵	Atherogenic index	377	0.82 ± 0.05	0.03943	1.0784	1.11783	0.03 ± 0.09

¹IUPAC Compendium of Chemical Terminology.

²SE Standard error.

³MUFA, PUFA and SFA were the total of all monounsaturated, polyunsaturated and saturated fatty acids, respectively.

⁴n-3 and n-6 were total of omega 3 and 6 fatty acids.

⁵Atherogenic index = [12:0 + 4(14:0) + 16:0]/(SFA + PUFA).

Table 2 Posterior means of variance components for IMF deposition and composition in Nellore by Bayes B

Trait	Genetic variance	Residual variance	Total variance	Genomic heritability
IMF (%)	0.16	0.48	0.64	0.25
C12:0 (mg/mg)	0.0001	0.0006	0.0007	0.18
C14:0	0.06	0.23	0.29	0.20
C14:1 cis-9	0.03	0.02	0.05	0.25
C15:0	0.006	0.04	0.046	0.12
C16:0	2.42	5.47	7.89	0.31
C16:1 cis-9	0.10	0.30	0.40	0.24
C17:0	0.004	0.02	0.024	0.17
C17:1	0.002	0.01	0.012	0.17
C18:0	1.23	5.28	6.51	0.19
C18:1 cis-9	6.08	7.11	13.19	0.46
C18:1 cis-11	0.04	0.33	0.37	0.11
C18:1 cis-12	0.005	0.03	0.035	0.13
C18:1, cis-13	0.02	0.002	0.022	0.09
C18:1 cis-15	0.0009	0.0004	0.00049	0.17
C18:1 trans-6, 7, 8	0.0008	0.005	0.0058	0.13
C18:1 trans-10, 11, 12	0.02	0.09	0.11	0.16
C18:1 trans-6	0.0003	0.002	0.0023	0.16
C18:2 cis 9 cis-12 n-6	0.03	0.23	0.26	0.13
C18:2 cis9, trans-11	0.0003	0.002	0.0023	0.12
C18:2 trans-11 cis-15	0.0001	0.0004	0.0005	0.22
C18:3 n-6	0.00007	0.0003	0.00037	0.21
C18:3 n-3	0.0003	0.002	0.0023	0.14
C20:1	0.0002	0.001	0.0012	0.16
C20:2	0.00005	0.00002	0.00007	0.22
C20:3 n-6	0.0003	0.002	0.0023	0.14
C20:4 n-6	0.002	0.02	0.022	0.08
C20:5 n-3	0.0002	0.001	0.0012	0.17
C22:5 n-3	0.0007	0.003	0.0037	0.16
C22:6 n-3	0.00006	0.0002	0.00026	0.24
SFA ¹	1.36	15.54	16.90	0.08
MUFA ¹	1.67	15.18	16.85	0.10
PUFA ¹	0.09	0.54	0.63	0.14
n-3 ²	0.004	0.013	0.017	0.25
n-6 ²	0.0005	0.003	0.0035	0.15
PUFA:SFA	0.003	0.02	0.023	0.11
n-6:n-3	0.66	1.23	1.89	0.34
AI ³	0.007	0.03	0.037	0.16

¹MUFA, PUFA and SFA were the total of all monounsaturated, polyunsaturated and saturated fatty acids, respectively.

²n-3 and n-6 were total of omega 3 and 6 fatty acids.

³Atherogenic index = $[12:0 + 4(14:0) + 16:0]/(SFA + PUFA)$.

35 traits were used for GWAS and these represented 23 different 1-Mb genomic regions (Table 3). These regions were distributed over 12 different chromosomes: 2, 3, 6, 7, 8, 9, 10, 11, 12, 17, 26 and 27 and the corresponding PCG

in these regions are reported in Table 3. Intramuscular fat was one of the traits that presented moderate heritability (0.25), however no region that explained more than 1% of the genetic variance for this trait was identified.

Table 3 QTL regions associated with fatty acid composition in Nellore steers by Bayes B

Group	Traits	QTL window (first and last SNP)	Number of SNP /window ¹	%Variance explained SNP window ¹	Chr	Map position (UMD 3.1 bovine assembly)	PCG ²
Saturated fatty acid	C14:0	rs43328164 - rs134696015	295	1.06	3	6009105 - 6999643	. ³
	C18:0	rs133274959 - rs136576856	213	3.46	3	25008982 - 25990012	<i>HMGCS2, PHGDH, HSD3B1, HAO2</i>
	C16:0	rs134160160 - rs109942510	285	1.38	3	26002777 - 26998679	<i>WARS2</i>
	C14:0	rs41627556 - rs110731616	332	1.43	9	36013595 - 36997263	<i>GNG11, RGS5</i>
	C18:0	rs136405986 - rs137105475	114	1.08	10	74001100 - 74957582	<i>DHRS7</i>
	C18:0	rs132804279 - rs109645596	102	2.08	11	105009792 - 105985714	<i>NUP214</i>
	C16:0	rs109773631 - rs135618512	277	1.53	12	13000697 - 13997298	-
	C12:0	rs42924061 - rs134942057	214	3.48	12	60000226 - 60994461	<i>SLITRK6</i>
Monounsaturated fatty acid	C14:1 cis-9	rs137683417 - rs29003360	232	1.86	2	26002605 - 26999718	<i>GAD1, Sp5</i>
	C18:1 cis-9	rs134160160 - rs109942510	285	1.40	3	26002777 - 26998679	<i>WARS2</i>
	C16:1 cis-9	rs133274959 - rs136576856	213	1.42	3	25008982 - 25990012	<i>HMGCS2, PHGDH, HSD3B1, HAO2</i>
	C18:1 cis-9	rs133274959 - rs136576856	213	2.57	3	25008982 - 25990012	<i>HMGCS2, PHGDH, HSD3B1, HAO2</i>
	C18:1 t-16	rs133728493 - rs109630757	264	2.56	6	50008629 - 50996673	-
	C16:1 cis-9	rs135892505 - rs109693564	184	1.04	7	80006438 - 80997126	-
	C16:1 cis-9	rs136405986 - rs137105475	114	1.18	10	74001100 - 74957582	<i>DHRS7</i>
	C14:1 cis-9	rs110517663 - rs137137362	127	1.43	11	27000446 - 27993515	<i>ABCG5</i>
	C18:1 cis-9	rs109773631 - rs135618512	277	1.91	12	13000697 - 13997298	-
	C14:1 cis-9	rs137754875 - rs135670282	203	1.55	12	66002783 - 66997356	<i>GPC6</i>
	C18:1 t-16	rs109872854 - rs134277482	250	2.17	17	41001273 - 41991683	<i>RAPGEF2, RPS27</i>
Polyunsaturated fatty acid	C22:5 n-3	rs132839318 - rs134264692	306	3.35	3	27001604 - 27997941	<i>SPAG7, WDR3</i>
	C18:2 t-11c-15	rs42404785 - rs133183089	144	1.69	3	72000121 - 72997801	<i>AQP7, LOXL2</i>
	C18:3 n-6	rs109612389 - rs133661384	205	1.34	7	34004443 - 34998434	-
	C18:2 t-11c-15	rs135377389 - rs133297940	267	3.49	8	68013821 - 68993778	-
	C22:5 n-3	rs110411459 - rs137802105	202	4.74	10	29000222 - 29962997	-

Table 3 QTL regions associated with fatty acid composition in Nellore steers by Bayes B (Continued)

	C20:5 n-3	rs137823965 - rs42507195	199	2.19	10	49005824 - 49980174	<i>RORA</i>
	C18:3 n-3	rs42582725 - rs133775322	132	1.33	17	24002089 - 24987264	-
	C18:2 t-11c-15	rs137602675 - rs133056879	187	1.01	26	20001875 - 20999270	-
	C20:3 n-6	rs134302284 - rs136338106	144	1.77	27	26000833 - 26975692	-
Total of MUFA	MUFA	rs133274959 - rs136576856	213	3.24	3	25008982 - 25990012	<i>HMGCS2</i> , <i>PHGDH</i> , <i>HSD3B1</i> , <i>HAO2</i>
		rs134160160 - rs109942510	285	1.13	3	26002777 - 26998679	<i>WARS2</i>
Total of n-3	n-3	rs110411459 - rs137802105	202	2.59	10	29000222 - 29962997	-
		rs137823965 - rs42507195	199	2.25	10	49005824 - 49980174	-
		rs1322839318 - rs134264692	306	1.37	3	27001604 - 27997941	<i>SPAG7</i> , <i>WDR3</i>
Total of n-6	n-6	rs134302284 - rs136338106	144	2.47	27	26000833 - 26975692	-

¹1 Mb window.

²Positional candidate genes.

³No putative candidate gene associated with the trait.

Saturated fatty acids

Eight genomic regions (1 Mb windows) explained more than 1% of genotypic variation for C12:0, C14:0, C16:0, and C18:0 (Table 3). These regions overlap with QTL previously reported for marbling score [28], backfat thickness [29], carcass weight and body weight in Angus cattle [30]. Additional file 1 shows in more detail the Manhattan plot of the proportion of genetic variance explained by window across the 29 autosomes for the important saturated fatty acids for beef palatability: C12:0 (lauric acid), palmitic acid (C16:0), stearic acid (C18:0).

BTA12 at 60 Mb QTL region was associated with C12:0 fatty acid and harbors the PCG *SLIT* and *NTRK*-like family, member 6 (*SLITRK6*). This gene is related to the lipopolysaccharide receptor complex (GO:0016021). Lipopolysaccharide (endotoxin) is an important component of gram-negative cell walls, which can start the inflammatory process by binding to the *SLITRK6* complex. This complex is located at the surface of innate immune cell, which presents an important role in obesity associated with insulin resistance [31].

BTA3 at 6 Mb and BTA9 at 36 Mb were associated with C14:0 fatty acid. In the first region, no PCG was identified. In the second region, the following PCGs were identified: guanine nucleotide binding protein (G protein), gamma 11 (*GNG11*) and regulator of G-protein signaling 5 (*RGSS5*). These genes are associated with G proteins that have been implicated in the regulation of body weight and metabolic function, hyperinsulinemia,

impaired glucose tolerance and resistance to insulin in mice [32].

BTA3 at 26 Mb and BTA12 at 13 Mb were associated with C16:0 fatty acid. In the first QTL region the tryptophanyl tRNA synthetase 2, mitochondrial (*WARS2*) gene was identified. In BTA12 at 13 Mb no PCG was identified. *WARS2* gene encodes an essential enzyme that catalyzes aminoacylation of tRNA with tryptophan. The *WARS2* protein contains a signal peptide for mitochondrial import, OMIM: 604733 [33], and is involved with regulation of fat distribution in human visceral and subcutaneous fat [34].

BTA3 at 25 Mb, BTA10 at 74 Mb and BTA11 at 105 Mb QTL regions were associated with C18:0 fatty acid. On BTA3 at 25 Mb four PCG were identified: 3-hydroxy-3-methylglutaryl-Coenzyme A synthase 2, mitochondrial (*HMGCS2*), phosphoglycerate dehydrogenase (*PHGDH*), hydroxy-delta-5-steroid dehydrogenase, 3 beta- and steroid delta-isomerase 1 (*HSD3B1*) and hydroxyacid oxidase 2, long chain (*HAO2*). *HMGCS2* was associated with fatty acid oxidation and ketogenesis in HepG2 cells [35] and is directly regulated by *PPAR-α* gene that is an important key regulator of β -oxidation. On the other hand the *PPAR-γ* expression induces lipogenesis by PXR activation in mice liver [36]. *PHGDH* is related to the regulation of gene expression according to gene ontology (GO) terms (GO:0010468), *HSD3B1* gene is associated with steroid biosynthesis (GO:0006694) and metabolic process (GO:0008202) and *HAO2* is associated with fatty acid oxidation (GO:0019395).

On BTA10 at 74 Mb QTL region, dehydrogenases/reductases (*SDRs*) (*DHRS7*) was identified. *DHRS7* catalyzes the oxidation/reduction of a wide range of substrates, including retinoid and steroids [37] and has high expression level in adipocyte and skeletal muscle [38]. In addition, this gene is responsible for the final step in cholesterol production, the conversion of 7-dehydrocholesterol to cholesterol [39].

On BTA11 at 105 Mb region, *NUP214* was identified. This gene is involved with intermembrane transport. The nucleoporins are characterized by phenylalanine-glycine rich (*FG*) repeat sequences and the *FG* domains have an unfolded structure and are responsible for interaction with importin-cargo complexes that move through the pore. One of these importin-cargo complexes is sterol regulatory element binding proteins-sterol-sensing accessory factor (*SREBP-SCAP*). This factor enters the nucleus, then binds to sterol regulatory elements (*SRE*) in the promoter regions of genes, whose products mediate the synthesis of cholesterol and fatty acids [40].

Monounsaturated fatty acids

Ten genomic regions (1 Mb region) explained more than 1% of genotypic variation for monounsaturated fatty acids, which relates C14:1 cis-9, C16:1 cis-9, C18:1 cis-9, and C18:1 trans-16 (Table 3). These regions overlap with QTL reported for carcass weight, marbling score in Angus [30], docosahexaenoic acid content in Charolais x Holstein crossbred cattle [41] and palmitoleic acid in dairy cattle [42]. Manhattan plots of the proportion of genetic variance explained by each 1-Mb window (2,527 windows) across the 29 autosomes for the most important fatty acid for beef quality and human health: myristoleic acid (C14:1 cis-9), palmitoleic acid (C16:1 cis-9), and oleic acid (C18:1 cis-9) and are in Figures 1, 2 and 3, respectively.

BTA2 at 26 Mb, BTA11 at 27 Mb, and BTA12 at 66 Mb were associated with C14:1 cis-9 fatty acid. On BTA2 at 26 Mb region, glutamate decarboxylase 1 (*GADI*) and specificity protein 5-transcription factor (*Sp5*) were identified. The *GADI* gene is involved with food behavior and insulin secretion. It was shown to be associated with morbid obesity [43] in humans, and body weight and daily gain in cattle [44]. *Sp5* transcription factors are involved in the regulation of pyruvate kinase, lactate dehydrogenase and fatty acid synthase in cancer cells [45,46]. On BTA11 at 27 Mb region, ATP-binding cassette, sub-family G (WHITE), member 5 (*ABCG5*) was identified. The *ABCG* family members were associated with cellular lipid-trafficking in macrophages and hepatocytes. This gene also presents an important role in the *PPAR γ* and *LXR* pathways [47]. On BTA12 at 66 Mb region, glypican 6 (*GPC6*) was identified. Glypicans family are involved with the control of cell growth and cell division. Other glypican gene, *GPC4* was associated with body fat distribution and obesity in humans

[48]. A previous GWAS using chicken population reported an association between *GPC6* and body weight, which has a stronger correlation with body fatness and obesity [49].

BTA3 at 25 Mb, BTA7 at 80 Mb and BTA10 at 74 Mb were associated with C16:1 cis-9 fatty acid. Both BTA3 at 25 Mb and BTA10 at 74 Mb regions were also associated with saturated fatty acids, where *HMGCS2*, *PHGDH*, *HSD3BI*, *HAO2*, and *DHRS7* were described above. On BTA7 at 80 Mb region, no PCG associated with this trait was identified.

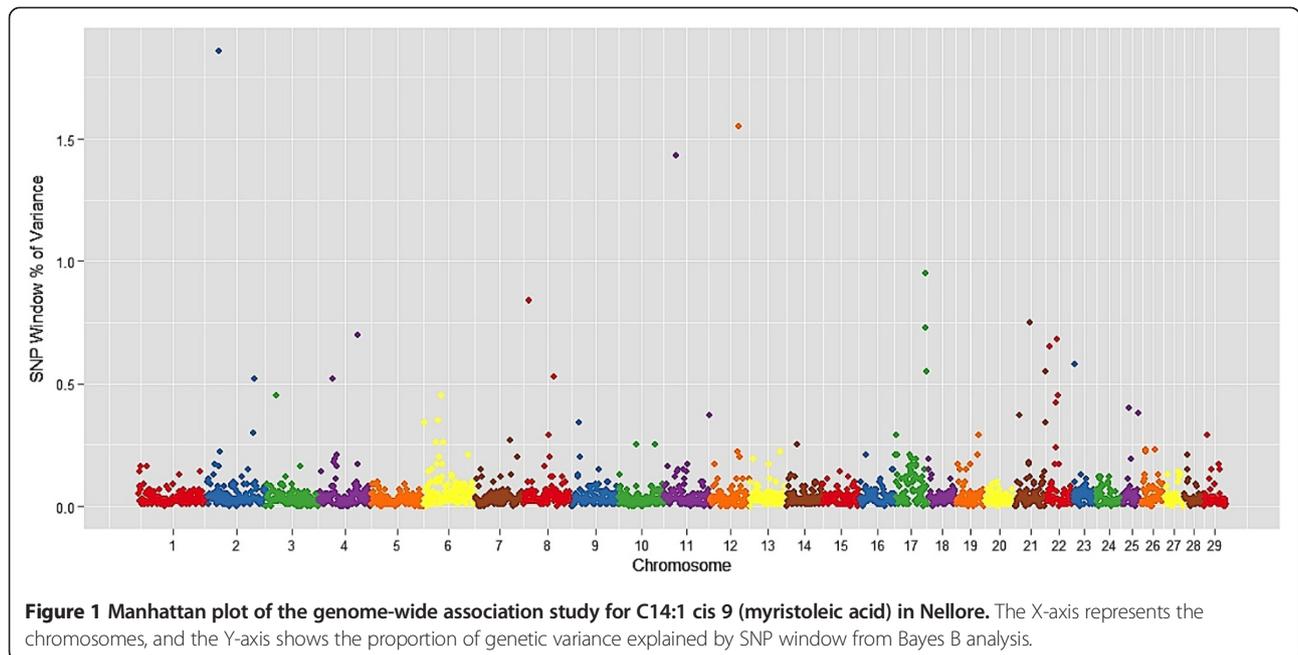
BTA3 at 25 Mb, BTA3 at 26 Mb, and BTA12 at 13 Mb were associated with C18:1 cis-9. The first two regions were also associated with saturated fatty acids (C18:0 and C16:0, respectively) and C16:1 cis-9. On BTA12 at 13 Mb region, there were no annotated genes [50].

BTA6 at 50 Mb and BTA17 at 41 Mb were associated with C18:1 trans-16. On BTA 6 at 50 Mb region, no PCG associated with this trait was identified, and on BTA17 at 41 Mb region one PCG was found: Rap guanine nucleotide exchange factor (*RAPGEF2*), that selectively and non-covalently interacts with diacylglycerol, a diester of glycerol and two fatty acids (GO:0019992).

Four genomic regions were associated with saturated and monounsaturated fatty acids. BTA3 at 26 Mb and BTA12 at 13 Mb were associated with C16:0 and C18:1 cis-9. BTA3 at 25 Mb was associated with C18:0 and C18:1 cis-9. BTA3 at 25 and BTA10 at 74 Mb were associated with C18:0 and C16:1 cis-9. We also observed a negative phenotypic correlation between C16:0 and C18:1 ($r = -0.65$) and between C18:0 and C18:1 ($r = -0.72$). Palmitic acid (C16:0) carbon chain skeleton is a predominant source for fatty acid elongation and desaturation, which might generate palmitoleic (C16:1 cis-9), stearic (C18:0), and oleic (C18:1 cis-9) fatty acids [51]. Examination of the 30 markers with the larger effect in regions of BTA3 at 26 Mb and BTA12 at 13 Mb (Table 4) and BTA3 at 25 Mb (Additional file 2) revealed that alleles positively associated with saturated fatty acid were negatively associated with unsaturated fatty acids. This indicates that QTL in these regions are directly or indirectly involved with elongation and/or desaturation.

Polyunsaturated fatty acids

Nine genomic regions (1 Mb window) explained > 1% of genotypic variation for group of polyunsaturated fatty acids, which relates C18:2 cis-9 cis-12 n-6, C18:2 trans-11 cis-15, C18:3 n-3, C18:3 n-6, C20:3 n-6, C20:5 n-3, and C22:5 n-3 (Table 3). These overlapped with QTL reported for marbling score, body weight in Angus cattle [30], intramuscular fat, saturated fatty acid content, stearic acid in Fleckvieh bulls [52] and RFI in half-sib families from Angus and Charolais [53]. Additional file 3 shows in more detail the Manhattan plot of the proportion of genetic variance explained by window across the 29 autosomes for the

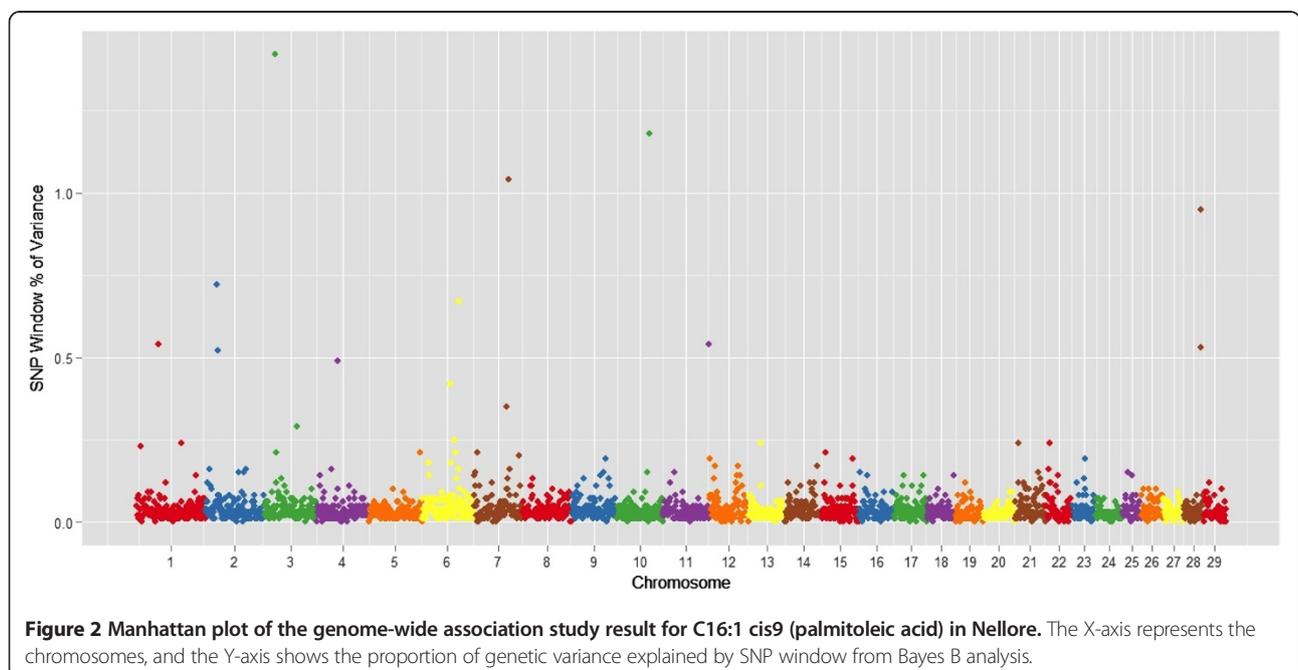


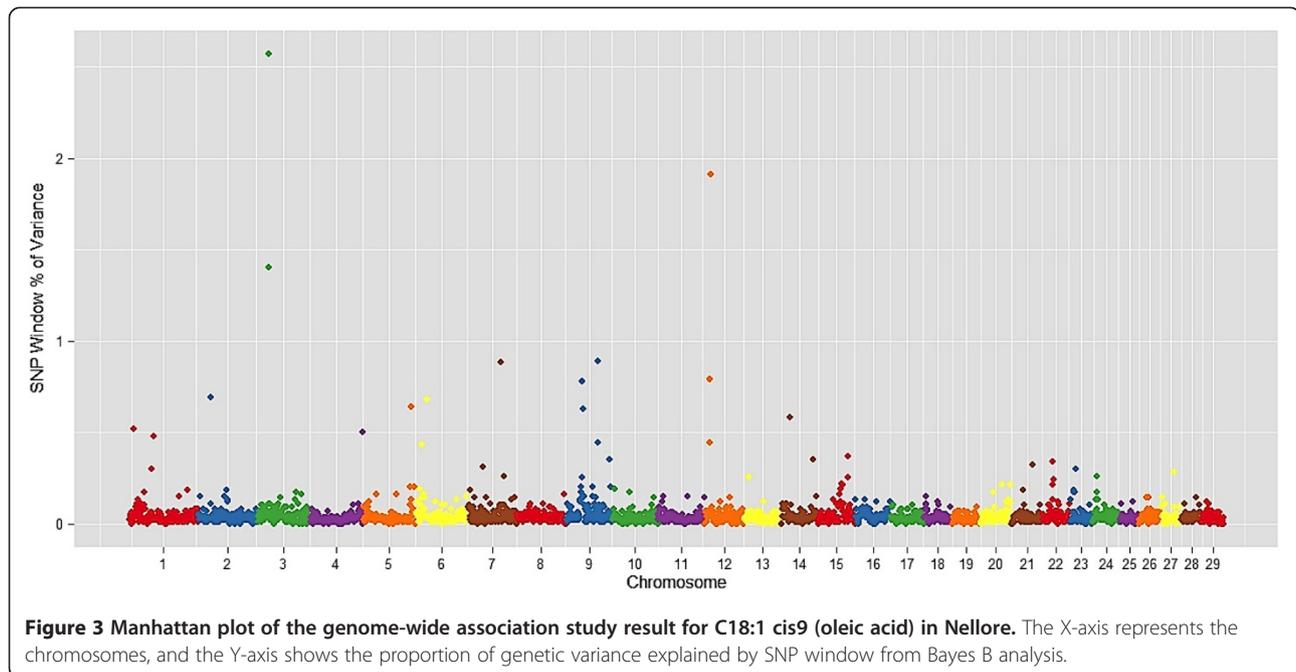
important fatty acid to human health: γ -linolenic acid (C18:3 n-3), α -linolenic acid (C18:3 n-6) and total of n-3 (n-3).

BTA3 at 72 Mb, BTA 8 at 68 Mb, and BTA26 at 20 Mb were associated with C18:2 trans-11 cis-15 fatty acid. On BTA3 at 72 Mb QTL region, two PCG related to lipid metabolism were observed: aquaporin 7 (*AQP7*) and lysil oxidase-like2 (*LOXL2*). *AQP7* is involved with the *PPAR*

signaling pathway [54], while *LOXL2* is associated with lean body mass in mouse [55]. On BTA 8 at 68 Mb and BTA26 at 20 Mb no PCG was associated with this trait.

BTA10 at 49 Mb region was associated with C20:5 n-3 fatty acid, which harbors the *RAR*-related orphan receptor A (*RORA*) gene that is related to steroid hormone receptor activity. When combined with a steroid hormone, it produces the signal within the





cell to initiate a change in cell activity or function (GO:0003707).

BTA3 at 27 Mb and BTA10 at 29 Mb QTL regions explained more than 1% of genetic variance for C22:3 n-3. The first region (BTA3 at 27 Mb) harbors two PCG: *SPAG17* and *WDR3*. These genes are involved with nucleus membrane (cellular components), the lipid bilayers that surround the nucleus and that form the nuclear envelope excluding the intermembrane space (GO:0031965). In the second region no PCG involved with this trait was identified. In other regions associated with C18:3 n-3 (BTA7 at 34 Mb), C18:3 n-6 (BTA17 at 24 Mb), and C20:3 n-6 (BTA27 at 26 Mb) no annotated genes were found [56].

The total of omega-3 (n-3) and omega-6 (n-6) GWAS result is consistent with individual polyunsaturated n-3 and n-6 fatty acids, where the same QTL regions were associated with these traits (BTA3 at 27 Mb and BTA 27 at 26 Mb).

GWAS have been used to investigate complex traits in many species including livestock [6]. Deposition and composition of fat in mammals are complex traits and are influenced by many loci throughout the genome [57]. GWAS provides one approach to understand the genetic variation in complex traits, by identifying regions that can be fine-mapped to identify individual loci responsible for variation [58]. In the present GWAS two interesting QTL regions BTA3 at 25 Mb and BTA3 at 26 Mb were associated with saturated, monounsaturated, and polyunsaturated fatty acids in *Bos indicus* cattle. These regions harbor interesting PCG, which are involved with lipid metabolism in different species.

Previous studies reported CCAAT/enhancer binding proteins (*C/EBPβ*), peroxisome proliferator-activated receptor gamma (*PPARγ* or *PPARG*), carnitine palmitoyltransferase-1 beta (*CPT-1β*), stearoylcoenzyme A desaturase (*SCD*), AMP-activated protein kinase alpha (*AMPKα*), and G-coupled protein receptor 43 (*GPR43*) genes to be related to fat deposition and fatty acid composition in Angus [59]. In this GWAS and in a recent GWAS publication using Angus cattle, the regions that harbor these genes were not associated with fat deposition and fatty acid composition [60]. However, the positional candidate genes *HMGCS2*, *PHGDH*, *HSD3B1* and *HAO2* identified in the present study are involved in the *PPAR* signaling pathway in human (PATH: hsa03320), carbon metabolism (PATH: bta01200), lipid metabolism and steroid hormone biosynthesis (PATH: bta00140), carbohydrate metabolism and glycosylate and decarboxylase metabolism (PATH: bta00630) according to Kegg: Kyoto Encyclopedia of Genes and Genomes [61], respectively.

This is the first GWAS for intramuscular fat deposition and composition in Nellore. GWAS results in *Bos taurus* (Angus and Japanese Black cattle) reported different genomic regions associated with fat deposition and composition than those reported herein [62,63]. Previously, it has been reported that a QTL on BTA19 was associated with fatty acid composition in *Bos taurus* breeds. This region on BTA19 harbors fatty acid synthase gene (*FASN*), which is an enzyme involved with *de novo* synthesis of long-chain fatty acid in mammalian, lipogenesis. *FASN* has been suggested as a candidate gene for fat traits in beef cattle [60,62,63].

Table 4 Top 30 markers effect in BTA3 at 26 Mb and BTA12 at 13 Mb QTL associated with C16:0 and C18:1 cis-9 in Nellore steers

BTA3 at 26 Mb				BTA12 at 13 Mb			
SNP name	Marker effect C16:0	Marker effect 18:1 cis-9	Position (Chr_Bp)	SNP name	Marker effect C16:0	Marker effect 18:1 cis-9	Position (Chr_bp)
rs136571109	1.49E-02	-8.42E-03	3_26426591	rs109310464	1.53E-02	-3.10E-02	12_13152910
rs110996601	1.45E-02	-6.58E-03	3_26425542	rs137546638	1.97E-03	-6.73E-03	12_13172924
rs134173038	1.40E-02	-6.05E-03	3_26427414	rs110182797	1.01E-03	-7.53E-04	12_13254936
rs137302352	4.10E-03	-2.29E-03	3_26400703	rs110951319	6.73E-04	-4.73E-04	12_13257070
rs135909241	9.74E-04	-2.55E-04	3_26446293	rs109014250	5.98E-04	-6.22E-04	12_13687406
rs109610305	7.91E-04	-1.72E-05	3_26497009	rs137052772	5.83E-04	-3.95E-04	12_13343491
rs137775131	5.12E-04	-7.50E-05	3_26501617	rs42625291	5.42E-04	-1.25E-03	12_13689345
rs109564367	5.01E-04	-8.52E-03	3_26073307	rs132782691	5.23E-04	-1.87E-04	12_13334837
rs110447932	4.30E-04	-8.35E-03	3_26072388	rs110645603	2.15E-04	-6.03E-05	12_13226418
rs109719131	3.84E-04	-9.90E-05	3_26465447	rs135746294	1.74E-04	-2.71E-05	12_13037523
rs110569793	3.62E-04	-9.64E-05	3_26476929	rs109497621	1.16E-04	-5.02E-05	12_13679288
rs133474163	3.39E-04	-1.56E-02	3_26143362	rs42625298	1.13E-04	-1.40E-04	12_13703328
rs110930607	3.22E-04	-1.90E-04	3_26332392	rs133583231	1.09E-04	-5.69E-05	12_13557660
rs134881809	3.10E-04	-2.36E-04	3_26505695	rs110721694	1.01E-04	-2.78E-03	12_13696926
rs111020805	2.70E-04	-1.92E-03	3_26108986	rs135841134	9.62E-05	-4.91E-05	12_13897680
rs134887465	2.68E-04	-2.04E-04	3_26385596	rs41667835	9.59E-05	-1.41E-04	12_13710147
rs110953593	2.61E-04	-8.47E-03	3_26075185	rs136432856	9.51E-05	3.08E-05	12_13493245
rs109658959	2.57E-04	-7.53E-03	3_26596824	rs133513235	9.45E-05	-4.21E-04	12_13251749
rs136570164	2.56E-04	-7.23E-05	3_26961906	rs137549822	8.60E-05	-6.65E-04	12_13081160
rs132696738	2.51E-04	-2.80E-04	3_26338744	rs110015470	8.45E-05	-3.18E-04	12_13682141
rs134559574	2.46E-04	-3.01E-05	3_26774155	rs133443666	8.30E-05	-1.46E-04	12_13520963
rs133573311	2.20E-04	-1.54E-04	3_26330114	rs208626835	8.10E-05	5.29E-06	12_13515819
rs135402139	1.94E-04	-1.26E-04	3_26954008	rs134142865	7.61E-05	-2.40E-04	12_13264826
rs135422840	1.71E-04	-3.00E-03	3_26020004	rs134240141	7.39E-05	-7.90E-05	12_13571494
rs136944072	1.66E-04	-2.63E-03	3_26149734	rs110659649	6.83E-05	-3.57E-05	12_13092209
rs110497471	1.56E-04	-2.50E-04	3_26576223	rs135136417	6.58E-05	-3.14E-05	12_13266029
rs137361087	1.45E-04	-3.15E-03	3_26144230	rs109064784	6.32E-05	-8.96E-05	12_13785143
rs110049045	1.37E-04	-7.04E-05	3_26975095	rs135831828	6.31E-05	-3.94E-05	12_13769919
rs133610187	1.36E-04	-2.31E-04	3_26290060	rs135423477	6.25E-05	-1.43E-04	12_13642422
rs110819650	1.35E-04	-4.69E-05	3_26979811	rs109147565	6.16E-05	-1.53E-03	12_13097096

Differences in SNP allele frequencies and linkage disequilibrium profile (LD between SNPs and causal variants) may explain the different marker effects between *Bos indicus* and *Bos taurus* cattle [64,65]. This explanation has been confirmed by Bolormaa and collaborators [66], who compared marker effects using GWAS for *Bos taurus* and *Bos indicus* animals, which demonstrated that a SNP effect depends on the origin of alleles and the QTL segregation. The QTL segregation could result from mutation lost or fixation of alleles in one of the breeds, and also that the mutations occurred after divergence of these breeds [67]. Furthermore, it is possible that the differences in physiological and metabolism factors could

contribute to the observed differences between different breeds [12].

Conclusion

The present study using BovineHD BeadChip (770 k) identified several 1-Mb SNP regions and genes within these regions that were associated with IMF and FA. The values of genomic heritabilities described in this study have not been reported before for Nellore cattle, and this information is important for breeding programs interesting in improving these traits. IMF deposition and composition are considered complex traits (polygenic) and are moderately heritable. In this

study it is apparent that IMF composition are affected by many loci with small effects. Identification of several genomic regions and putative positional genes associated with lipid metabolism reported here should contribute to the knowledge of the genetic basis of IMF and FA deposition and composition in Nellore cattle (*Bos indicus* breed) and lead to selection for those traits to improve human nutrition and health.

Methods

Animals were handled and managed according to Institutional Animal Care and Use Committee Guidelines from Brazilian Agricultural Research Corporation – EMBRAPA approved by the president, Dr. Rui Machado.

Animals and phenotypes

Nellore steers (386) bred in the Brazilian Agricultural Research Corporation (EMBRAPA/Brazil) experimental breeding herd between 2009 and 2011 were available for this study. Steers were sired by 34 unrelated sires, and were selected to represent the main genealogies used in Brazil according to the National Summary of Nellore produced by the Brazilian Association of Zebu Breeders (ABCZ) and National Research Center for Beef Cattle. Animals were raised in feedlots under identical nutrition and handling conditions until slaughter at an average age of 25 months [68]. Steaks (2.54 cm thick) from the *Longissimus dorsi* muscle between the 12th and 13th ribs were collected 24 hours after slaughter.

Muscle samples (~100 g) were lyophilized and ground for IMF and FA analysis. The IMF was obtained using an Ankom XT20 extractor as described [69]. FA analysis was conducted as described by Hara and Radin [70], except the hexane to propanol ratio was increased to 3:2. Approximately 4 g of LD muscle was lyophilized, ground in liquid nitrogen, mixed with 28 mL of hexane/propanol (3:2 vol/vol) and homogenized for 1 min. Samples were vacuum filtered and 12 ml sodium sulfate (67 mg mL⁻¹) solution was added and agitated for 30 s. The supernatant was transferred to a tube with 2 g of sodium sulfate and insufflated with N₂, after which the tube was sealed and incubated at room temperature for 30 min. Subsequently, the liquid was transferred to 10 mL test tube, insufflated with N₂, sealed and kept at -20°C until dry with N₂ for methylation. The extracted lipids were hydrolyzed and methylated as described by Christie [71], except that hexane and methyl acetate were used instead of hexane:diethyl ether:formic acid (90:10:1). Around 40 mg of lipids were transferred to a tube containing 2 mL of hexane. Subsequently, 40 µL of methyl acetate were added, the sample agitated, and 40 µL of methylation solution (1.75 mL of methanol/0.4 mL of 5.4 mol/L of sodium metoxyde) were added. This mixture was agitated for

2 min and incubated for 10 min at room temperature. Then 60 µL of finishing solution (1 g of oxalic acid/30 mL of diethyl ether) were added and the mixture was agitated for 30 s, after which 200 mg of calcium chloride was added. The sample was then mixed and incubated at room temperature for 1 h. Samples were centrifuged at 3200 rpm, for 5 min at 5°C. The supernatant was collected for determination of fatty acids. Fatty acid methyl esters were quantified with a gas chromatograph (ThermoFinnigan, Termo Electron Corp., MA, USA) equipped with a flame ionization detector and a 100 m Supelco SP-2560 (Supelco Inc., PA, USA) fused silica capillary column (100 m, 0.25 mm and 0.2 µm film thickness). The column oven temperature was held at 70°C for 4 min, then increased to 170°C at a rate of 13°C min⁻¹, and subsequently increased to 250°C at a rate of 35°C min⁻¹, and held at 250°C for 5 min. The gas fluxes were 1.8 mL min⁻¹ for carrier gas (He), 45 mL min⁻¹ for make-up gas (N₂), 40 mL min⁻¹ for hydrogen, and 450 mL min⁻¹ for synthetic flame gas. One µL sample was analyzed. Injector and detector temperatures were 250 and 300°C, respectively. Fatty acids were identified by comparison of retention time of methyl esters of the samples with standards of fatty acids butter reference BCR-CRM 164, Anhydrous Milk Fat-Producer (BCR Institute for Materials and Reference Measurements) and also with commercial standard for 37 fatty acids Supelco TM Component FAME Mix (cat 18919, Supelco, Bellefonte, PA). The nomenclature of fatty acids follow IUPAC Compendium [72]. Fatty acids were quantified by normalizing the area under the curve of methyl esters using Chromquest 4.1 software (Thermo Electron, Italy). Fatty acids were expressed as a weight percentage (mg/mg). These analyses were performed at the Animal Nutrition and Growth Laboratory at ESALQ, Piracicaba, São Paulo, Brazil.

DNA extraction and genotypic data

DNA was isolated from blood as described by Tizioto et al. [68]. Genotyping was performed at the Bovine Functional Genomics Laboratory ARS/USDA and Genomics Center at ESALQ, Piracicaba, São Paulo, Brazil using BovineHD 770 k BeadChip (Infinium BeadChip, Illumina, San Diego, CA) according to manufacturer's protocol. Genotypes were obtained in Illumina A/B allele format and used to represent a covariate value at each locus coded as 0, 1, or 2, representing the number of B alleles. Missing genotypes, represented < 0.2% of genotypes and were replaced with the average covariate value at that locus. Initial visualization and data analysis was performed by GenomeStudio Data Analysis Software [73]. The SNPs with call rate ≤ 95%, minor allele frequency (MAF) ≤ 5%, those located on sex chromosomes and those not mapped in the *Bos taurus* UMD 3.1 assembly were removed. After filtering, a total of 449,363 SNP were utilized in GWAS.

Descriptive statistics and heritability

Descriptive statistics for IMF and FA were estimated using PROC MEANS and normality tests were performed using PROC UNIVARIATE in SAS (Ver. 9.3; SAS Inst. Cary, NC). SAS PROC MIXED was used to test independent sources for significance. Fixed effects included contemporary group classes (animals with the same origin, birth year and slaughter date) and hot carcass weight as a covariate. Animal and residuals were fitted as random effects. Restricted maximum likelihood was used to estimate animal and residual variance components, heritability and standard error (SE) using ASREML software [74]. The model used in single-trait analyses of all traits was, $y = Xb + Zu + e$, where y is the vector of observations representing the trait of interest (dependent variable), X and Z are the design or incidence matrices for the vectors of fixed and random effects in b and u , respectively, and e was the vector of random residuals. The variance of vector u was $G\sigma_m^2$ for the genomic analyses where G is the genomic relationship matrix derived from SNP markers using allele frequencies as suggested by VanRaden [75], with σ_m^2 being the marker-based additive genetic variance.

Genome wide association study

Associations between SNP and phenotypes (IMF and FA) were obtained using Bayes B, which analyzed all SNP data simultaneously and assumed a different genetic variance for each SNP locus [76,77]. The prior genetic and residual variances were estimated using Bayes C [78], with π being 0.9997. The model equation was:

$$y = Xb + \sum_{j=1}^k a_j \beta_j \delta_j + e,$$

where y was the vector of the phenotypic values, X was the incidence matrix for fixed effects, b was the vector of fixed effects defined above, k was the number of SNP loci (449,363), a_j was the column vector representing the SNP covariate at locus j coded as the number of B alleles, β_j was the random substitution effect for locus j , which conditional on σ_β^2 was assumed to be normally distributed $N(0, \sigma_\beta^2)$ when $\delta_j = 1$ but $\beta_j = 0$ when $\delta_j = 0$, with δ_j being a random 0/1 variable indicating the absence (with probability π) or presence (with probability $1-\pi$) of locus j in the model, and e was the vector of the random residual effects assumed normally distributed $N(0, \sigma_e^2)$. The variance σ_β^2 (or σ_e^2) was a *priori* assumed to follow a scaled inverse Chi-square with $\nu_\beta = 4$ (or $\nu_e = 10$) degrees of freedom and scale parameter S_β^2 (or S_e^2). The scale parameter for markers was derived as a function of the assumed known genetic variance of the population, based on the average SNP allele frequency and number of SNP assumed to have nonzero effects

based on parameter π being 0.9997. This procedure used GenSel software [8] to obtain the posterior distributions of SNP effects using Markov chain Monte Carlo (MCMC). This comprised a burn-in period of 1,000 iterations from which results were discarded, followed by 40,000 iterations from which results were accumulated to obtain the posterior mean effect of each SNP. In the Bayesian variable selection multiple-regression models with $\pi = 0.9997$ about 100-150 SNP markers were fitted simultaneously in each MCMC iteration. Inference of associations in these multiple-regression models was based on 1-Mb genomic windows rather than on single markers [8,79]. Genomic windows were constructed from the chromosome and base-pair positions denoted in the marker map file [8] based on UMD3.1 bovine assembly.

The SNP effects from every 40th post burn in iteration were used to obtain samples from the posterior distribution of the proportion of variance accounted for by each window from 1,000 MCMC samples of genomic merit for each animal following Onteru et al. [79] and Peters et al. [7]. In the present study there were 2,527 1 Mb SNP windows across the 29 autosomes. The proportion of genetic variance explained by each window in any particular iteration was obtained by dividing the variance of window BV by the variance of whole genome BV in that iteration. The window BV was computed by multiplying the number of alleles that represent the SNP covariates for each consecutive SNP in a window by their sampled substitution effects in that iteration.

All traits were used for GWAS (Additional file 4) and genomic heritability estimate, but only the ones with genomic heritability ≥ 0.10 were reported. Fatty acids were indexed as groups of saturated, monounsaturated, polyunsaturated fatty acid, total of saturated fatty acid (SFA), total monounsaturated (MUFA), total of polyunsaturated (PUFA), total of omega 3 (n-3) and total of omega 6 (n-6). Genome windows with the highest posterior mean proportion of genetic variance $\geq 1\%$ were considered the most important regions associated with the traits, and were declared the most promising QTL regions.

Positional candidate genes were investigated for 1 Mb windows using the Cattle Genome Browser [50] and UCSC Genome Browser [80], which allowed visualization of SNP based on *Bos taurus* genome assembly UMD 3.1. Animal QTL database (Animal QTLdb) was used to search for published QTL and trait mapping data [56]. Gene annotations were retrieved from Ensembl Genes 71 Database using Biomart software [54,81]. The functional classification of genes was done using DAVID [54] and BioGPS [38] online annotation databases. Those genes reported to be involved in fatty acid and lipid metabolism were selected as positional candidate genes.

Availability of supporting data

The data sets supporting the results of this article are included within the article and its additional files.

Additional files

Additional file 1: Manhattan plot of the genome-wide association study result for A) C12:0 (lauric acid) B) C16:0 (palmitic acid) C) C18:0 (stearic acid) in Nellore. The X-axis represents the chromosomes, and the Y-axis shows the proportion of genetic variance explained by SNP window from Bayes B analysis.

Additional file 2: Top 30 markers effect in BTA3 at 25 Mb associated with C18:0 and C18:1 cis-9 in Nellore steers.

Additional file 3: Manhattan plot of the genome-wide association study result for A) C18:3 n-3 (α -linolenic acid) B) C18:3 n-6 (α -linolenic acid) C) n-3 (total of n-3) in Nellore. The X-axis represents the chromosomes, and the Y-axis shows the proportion of genetic variance explained by SNP window from Bayes B analysis.

Additional file 4: Top three QTL regions associated with IMF deposition and composition traits in Nellore by Bayes B.

Abbreviations

IMF: Intramuscular fat; LDL: Low density lipoprotein; MUFA: Total of monounsaturated fatty acid; FA: Fatty acid; SNP: Single nucleotide polymorphism; GWAS: Genome-wide association study; QTL: Quantitative trait loci; SFA: Saturated fatty acid; PUFA: Total of polyunsaturated fatty acid; CLA: Conjugated linoleic acid; PCG: Putative candidate gene; GO: Gene ontology; MAF: Minor allele frequency; SE: Standard error; MCMC: Markov chain Monte Carlo; SFA: Total of saturated fatty acid; n-3: Total of omega 3; n-6: Total of omega 6.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

ASMC, LCAR, LLC, RRT, GBM, DPDL, RTN and MMA conceived and designed the experiment; ASMC, LCAR, LLC, RTN, RRT, MLN, ASC and TSS performed the experiments; ASMC, LCAR, MAM, PSNO, DJG, JMR and LLC did analysis and interpretation of results; ASMC, LCAR, DJG, JMR and LLC drafted the manuscript. All authors read and approved the final manuscript.

Acknowledgements

This study was conducted with funding from EMBRAPA (Macroprograma 1, 01/2005) and FAPESP (process number 2011/00005-7 and 2012/02383-1). LR, MA, GBM and LC were granted CNPq fellowships. The authors would like to acknowledge the collaborative efforts among EMBRAPA, University of São Paulo, Iowa State University and USDA ARS Bovine Functional Genomics Laboratory, and fruitful discussions with Madhi Saatchi, Xiaochen Sun and Anna Wolc.

Author details

¹Department of Animal Science, University of São Paulo, Piracicaba SP 13418-900, Brazil. ²Embrapa Southeast-Cattle Research Center, São Carlos, SP 13560-970, Brazil. ³Department of Genetics and Evolution, Federal University of São Carlos, São Carlos, SP 13565-905, Brazil. ⁴United States Department of Agriculture, Agricultural Research Service, Bovine Functional Genomics Laboratory, Beltsville, Maryland 20705, USA. ⁵Department of Animal Science, Iowa State University, Ames, IA 50011, USA.

Received: 16 October 2013 Accepted: 28 February 2014

Published: 25 March 2014

References

1. Laborde FL, Mandell IB, Tosh JJ, Wilton JW, Buchanan-Smith JG: **Breed effects on growth performance, carcass characteristics, fatty acid composition, and palatability attributes in finishing steers.** *J Anim Sci* 2001, **79**(2):355–365.
2. McNeill S, van Elswyk ME: **Red meat in global nutrition.** In *Meat Sci*, Volume 92. England: 2012 Elsevier Ltd; 2012:166–173.
3. Jakobsen MU, Overvad K, Dyerberg J, Heitmann BL: **Intake of ruminant trans fatty acids and risk of coronary heart disease.** *Int J Epidemiol* 2008, **37**(1):173–182.
4. Rule DC, MacNeil MD, Short RE: **Influence of sire growth potential, time on feed, and growing-finishing strategy on cholesterol and fatty acids of the ground carcass and longissimus muscle of beef steers.** *J Anim Sci* 1997, **75**(6):1525–1533.
5. Matukumalli LK, Lawley CT, Schnabel RD, Taylor JF, Allan MF, Heaton MP, O'Connell J, Moore SS, Smith TP, Sonstegard TS, Van Tassel CP: **Development and characterization of a high density SNP genotyping assay for cattle.** *PLoS One* 2009, **4**(4):e5350.
6. Hayes BJ, Bowman PJ, Chamberlain AC, Verbyla K, Goddard ME: **Accuracy of genomic breeding values in multi-breed dairy cattle populations.** *Genet Sel Evol* 2009, **41**:51.
7. Peters SO, Kizilkaya K, Garrick DJ, Fernando RL, Reedy JM, Weaver RL, Silver GA, Thomas MG: **Bayesian genome-wide association analysis of growth and yearling ultrasound measures of carcass traits in Brangus heifers.** *J Anim Sci* 2012, **90**(10):3398–3409.
8. Garrick DJ, Fernando RL: **Implementing a QTL detection study (GWAS) using genomic prediction methodology.** *Methods Mol Biol* 2013, **1019**:275–298.
9. Kizilkaya K, Tait RG, Garrick DJ, Fernando RL, Reedy JM: **Genome-wide association study of Infectious Bovine Keratoconjunctivitis in Angus cattle.** *BMC Genet* 2013, **14**:23.
10. Prakash A, Stigler M: *FAO Statistical Yearbook 2012: WORLD FOOD AND AGRICULTURE.* Rome, Italy: Food and Agriculture Organization of the United Nations; 2012.
11. Picard B, Lefaucheur L, Berri C, Duclos MJ: **Muscle fibre ontogenesis in farm animal species.** *Reprod Nutr Dev* 2002, **42**(5):415–431.
12. Lehnert SA, Reverter A, Byrne KA, Wang Y, Natrass GS, Hudson NJ, Greenwood PL: **Gene expression studies of developing bovine longissimus muscle from two different beef cattle breeds.** *BMC Dev Biol* 2007, **7**:95.
13. Duarte MS, Paulino PV, Das AK, Wei S, Seroa NV, Fu X, Harris SM, Dodson MV, Du M: **Enhancement of adipogenesis and fibrogenesis in skeletal muscle of Wagyu compared with Angus cattle.** *J Anim Sci* 2013, **91**(6):2938–2946.
14. Du M, Wang X, Das AK, Yang Q, Duarte MS, Dodson MV, Zhu M: **Manipulating mesenchymal progenitor cell differentiation to optimize performance and carcass value of beef cattle.** *J Anim Sci* 2013, **91**(3):1419–1427.
15. Huerta-Leidenz NO, Cross HR, Savell JW, Lunt DK, Baker JF, Smith SB: **Fatty acid composition of subcutaneous adipose tissue from male calves at different stages of growth.** *J Anim Sci* 1996, **74**(6):1256–1264.
16. Holloway JW, Savell JW, Hamby PL, Baker JF, Stouffer JR: **Relationships of empty-body composition and fat distribution to live animal and carcass measurements in Bos indicus-Bos taurus crossbred cows.** *J Anim Sci* 1990, **68**(7):1818–1826.
17. Charles DD, Johnson ER: **Breed differences in amount and distribution of bovine carcass dissectable fat.** *J Anim Sci* 1976, **42**:332.
18. Nuernberg K, Dannenberger D, Nuernberg G, Ender K, Voigt J, Scollan ND, Wood JD, Nute GR, Richardson RT: **Effect of grass-based and a concentrate feeding system on meat quality characteristics and fatty acid composition of longissimus muscle in different cattle breeds.** *Livest Prod Sci* 2005, **94**:137–147.
19. Padre R, Aricetti JA, Moreira FB, Mizubuti IY, Do Prado IN, Visentainer JV, de Souza NE, Matsushita M: **Fatty acid profile, and chemical composition of Longissimus muscle of bovine steers and bulls finished in pasture system.** *Meat Sci* 2006, **74**(2):242–248.
20. Bressan MC, Rossato LV, Rodrigues EC, Alves SP, Bessa RJ, Ramos EM, Gama LT: **Genotype x environment interactions for fatty acid profiles in Bos indicus and Bos taurus finished on pasture or grain.** *J Anim Sci* 2011, **89**(1):221–232.
21. Prado IN, Moreira FB, Matsushita M, de Souza NE: **Longissimus dorsi fatty acids composition of Bos indicus and Bos indicus x Bos taurus crossbred steers finished in pasture.** *Braz Arch Biol Technol* 2003, **46**(4):601–608.
22. USDA: *National Nutrient Database for Standard Reference, Release 23 (SR 23) [database on the Internet].* Washington, DC: USDA, Agricultural Research Service; 2010. Available from: http://www.ars.usda.gov/SP2UserFiles/Place/12354500/Data/SR23/sr23_doc.pdf.
23. Ruiz M, Matsushita M, Visentainer J, Hernandez J, Ribeiro E, Shimokomaki M, Reeves J, Souza N: **Proximate chemical composition and fatty acid**

- profiles of Longissimus thoracis from pasture fed LHRH immunocastrated, castrated and intact Bos indicus bulls. *S Afr J Anim Sci* 2005, **35**(1):13–18.
24. Simopoulos AP: **The importance of the ratio of omega-6/omega-3 essential fatty acids.** *Biomed Pharmacother* 2002, **56**(8):365–379.
25. Simopoulos AP: **The omega-6/omega-3 fatty acid ratio, genetic variation, and cardiovascular disease.** *Asia Pac J Clin Nutr* 2008, **17**(Suppl 1):131–134.
26. Tait R Jr, Zhang S, Knight T, Minick Bormann J, Beitz D, RJ M: **Heritability estimates for fatty acid quantity in Angus beef.** *J Anim Sci* 2007, **85**(Suppl. 2):58. Abstr.
27. Nogi T, Honda T, Mukai F, Okagaki T, Oyama K: **Heritabilities and genetic correlations of fatty acid compositions in longissimus muscle lipid with carcass traits in Japanese Black cattle.** *J Anim Sci* 2011, **89**(3):615–621.
28. Casas E, Stone RT, Keele JW, Shackelford SD, Kappes SM, Koohmaraie M: **A comprehensive search for quantitative trait loci affecting growth and carcass composition of cattle segregating alternative forms of the myostatin gene.** *J Anim Sci* 2001, **79**(4):854–860.
29. Casas E, Shackelford SD, Keele JW, Koohmaraie M, Smith TP, Stone RT: **Detection of quantitative trait loci for growth and carcass composition in cattle.** *J Anim Sci* 2003, **81**(12):2976–2983.
30. McClure MC, Morsci NS, Schnabel RD, Kim JW, Yao P, Rolf MM, McKay SD, Gregg SJ, Chapple RH, Northcutt SL, Taylor JF: **A genome scan for quantitative trait loci influencing carcass, post-natal growth and reproductive traits in commercial Angus cattle.** In *Anim Genet*, Volume 41. England: 2010 The Authors, Animal Genetics 2010 Stichting International Foundation for Animal Genetics; 2010:597–607.
31. Shi H, Kokoeva MV, Inouye K, Tzamelis I, Yin H, Flier JS: **TLR4 links innate immunity and fatty acid-induced insulin resistance.** *J Clin Invest* 2006, **116**(11):3015–3025.
32. Deng W, Wang X, Xiao J, Chen K, Zhou H, Shen D, Li HT, Qizhu: **Loss of Regulator of G Protein Signaling 5 Exacerbates Obesity, Hepatic Steatosis.** *Inflamm Insulin Resist* 2012, **7**(1):e30256.
33. McKusick VA: **Mendelian Inheritance in Man and its online version, OMIM.** *Am J Hum Genet* 2007, **80**(4):588–604.
34. Schleinitz D, Kloting N, Lindgren CM, Breitfeld J, Dietrich A, Schon MR, Lohmann T, Dressler M, Stumvoll M, McCarthy MI, Blüher M, Kovacs P: **Fat depot-specific mRNA expression of novel loci associated with waist-hip ratio.** *Int J Obes (Lond)* 2014, **38**(1):120–125.
35. Vila-Brau A, de Sousa-Coelho AL, Mayordomo C, Haro D, Marrero PF: **Human HMGCS2 regulates mitochondrial fatty acid oxidation and FGF21 expression in HepG2 cell line.** *J Biol Chem* 2011, **286**(23):20423–20430.
36. Wada T, Gao J, Xie W: **PXR and CAR in energy metabolism.** *Trends Endocrinol Metab* 2009, **20**(6):273–279.
37. Haeseleer F, Palczewski K: **Short-chain dehydrogenases/reductases in retina.** *Methods Enzymol* 2000, **316**:372–383.
38. Wu C, Orozco C, Boyer J, Leglise M, Goodale J, Batalov S, Hodge CL, Haase J, Janes J, Huss JW 3rd, Su AI: **BioGPS: an extensible and customizable portal for querying and organizing gene annotation resources.** *Genome Biol* 2009, **10**(11):R130.
39. Porter FD: **RSH/Smith-Lemli-Opitz syndrome: a multiple congenital anomaly/mental retardation syndrome due to an inborn error of cholesterol biosynthesis.** In *Mol Genet Metab*, Volume 71. United States: 2000 Academic Press; 2000:163–174.
40. Zhou T, Li S, Zhong W, Vihervaara T, Beaslas O, Perttila J, Luo W, Jiang Y, Lehto M, Oikkonen VM, Yan D: **OSBP-related protein 8 (ORP8) regulates plasma and liver tissue lipid levels and interacts with the nucleoporin Nup62.** *PLoS One* 2011, **6**(6):e21078.
41. Gutierrez-Gil B, Williams JL, Homer D, Burton D, Haley CS, Wiener P: **Search for quantitative trait loci affecting growth and carcass traits in a cross population of beef and dairy cattle.** *J Anim Sci* 2009, **87**(1):24–36.
42. Morris CA, Cullen NG, Glass BC, Hyndman DL, Manley TR, Hickey SM, McEwan JC, Pitchford WS, Bottema CD, Lee MA: **Fatty acid synthase effects on bovine adipose fat and milk fat.** *Mamm Genome* 2007, **18**(1):64–74.
43. Meyre D, Boutin P, Tounian P, Deweilder M, Aout M, Jouret B, Heude B, Weill J, Tauber M, Tounian P, Froquel P: **Is glutamate decarboxylase 2 (GAD2) a genetic link between low birth weight and subsequent development of obesity in children?** *J Clin Endocrinol Metab* 2005, **90**(4):2384–2390.
44. Li C, Aldai N, Vinsky M, Dugan ME, McAllister TA: **Association analyses of single nucleotide polymorphisms in bovine stearoyl-CoA desaturase and fatty acid synthase genes with fatty acid composition in commercial cross-bred beef steers.** *Anim Genet* 2012, **43**(1):93–97.
45. Black AR, Black JD, Azizkhan-Clifford J: **Sp1 and kruppel-like factor family of transcription factors in cell growth regulation and cancer.** In *J Cell Physiol*, Volume 188. United States: 2001 Wiley-Liss, Inc; 2001:143–160.
46. Archer MC: **Role of sp transcription factors in the regulation of cancer cell metabolism.** *Genes Cancer* 2011, **2**(7):712–719.
47. Schmitz G, Langmann T, Heimerl S: **Role of ABCG1 and other ABCG family members in lipid metabolism.** *J Lipid Res* 2001, **42**(10):1513–1520.
48. Gesta S, Bluher M, Yamamoto Y, Norris AW, Berndt J, Kralisch S, Boucher J, Lewis C, Kahn CR: **Evidence for a role of developmental genes in the origin of obesity and body fat distribution.** *Proc Natl Acad Sci U S A* 2006, **103**(17):6676–6681.
49. Gu X, Feng C, Ma L, Song C, Wang Y, Da Y, Li H, Chen K, Ye S, Ge C, Hu X, Li N: **Genome-wide association study of body weight in chicken F2 resource population.** *PLoS One* 2011, **6**(7):e21872.
50. **Cattle Genome UMD3.1.** [http://www.animalgenome.org/cgi-bin/gbrowse/bovine/]
51. Inagaki K, Aki T, Fukuda Y, Kawamoto S, Shigeta S, Ono K, Suzuki O: **Identification and expression of a rat fatty acid elongase involved in the biosynthesis of C18 fatty acids.** *Biosci Biotechnol Biochem* 2002, **66**(3):613–621.
52. Barton L, Kott T, Bures D, Rehak D, Zahradkova R, Kottova B: **The polymorphisms of stearoyl-CoA desaturase (SCD1) and sterol regulatory element binding protein-1 (SREBP-1) genes and their association with the fatty acid profile of muscle and subcutaneous fat in Fleckvieh bulls.** In *Meat Sci*, Volume 85. England: 2009 Elsevier Ltd; 2010:15–20.
53. Sherman EL, Nkrumah JD, Li C, Bartusiak R, Murdoch B, Moore SS: **Fine mapping quantitative trait loci for feed intake and feed efficiency in beef cattle.** *J Anim Sci* 2009, **87**(1):37–45.
54. da Huang W, Sherman BT, Lempicki RA: **Bioinformatics enrichment tools: paths toward the comprehensive functional analysis of large gene lists.** *Nucleic Acids Res* 2009, **37**(1):1–13.
55. Baker EJ, Jay JJ, Bubier JA, Langston MA, Chesler EJ: **GeneWeaver: a web-based system for integrative functional genomics.** *Nucleic Acids Res* 2012, **40**(Database issue):D1067–D1076.
56. Hu ZL, Park CA, Wu XL, Reedy JM: **Animal QTLdb: an improved database tool for livestock animal QTL/association data dissemination in the post-genome era.** *Nucleic Acids Res* 2013, **41**(Database issue):D871–D879.
57. van Laere AS, Nguyen M, Braunschweig M, Nezer C, Collette C, Moreau L, Archibald AL, Haley CS, Buys N, Tally M, Andersson G, Georges M, Andersson L: **A regulatory mutation in IGF2 causes a major QTL effect on muscle growth in the pig.** *Nature* 2003, **425**(6960):832–836.
58. Kemper KE, Goddard ME: **Understanding and predicting complex traits: knowledge from cattle.** *Hum Mol Genet* 2012, **21**(R1):R45–51.
59. Smith SB, Go GW, Johnson BJ, Chung KY, Choi SH, Sawyer JE, Silvey DT, Gilmore LA, Ghahramany G, Kim KH: **Adipogenic gene expression and fatty acid composition in subcutaneous adipose tissue depots of Angus steers between 9 and 16 months of age.** *J Anim Sci* 2012, **90**(8):2505–2514.
60. Saatchi M, Garrick DJ, Tait RG Jr, Mayes MS, Drewnoski M, Schoonmaker J, Diaz C, Beitz DC, Reedy JM: **Genome-wide association and prediction of direct genomic breeding values for composition of fatty acids in Angus beef cattle.** *BMC Genomics* 2013, **14**(1):730.
61. Kanehisa M, Goto S, Sato Y, Furumichi M, Tanabe M: **KEGG for integration and interpretation of large-scale molecular data sets.** *Nucleic Acids Res* 2012, **40**(Database issue):D109–D114.
62. Uemoto Y, Abe T, Tameoka N, Hasebe H, Inoue K, Nakajima H, Shoji N, Kobayashi M, Kobayashi E: **Whole-genome association study for fatty acid composition of oleic acid in Japanese Black cattle.** *Anim Genet* 2010, **42**(2):141–148.
63. Ishii A, Yamaji K, Uemoto Y, Sasago N, Kobayashi E, Kobayashi N, Matsuhashi T, Maruyama S, Matsumoto H, Sasazaki S, Mannen H: **Genome-wide association study for fatty acid composition in Japanese Black cattle.** *Anim Sci J* 2013, **84**(10):675–682.
64. Tizioto PC, Decker JE, Taylor JF, Schnabel RD, Mudadu MA, Silva FL, Mourão GB, Coutinho LL, Tholon P, Sonstegard TS, Rosa AN, Alencar MM, Tullio RR, Medeiros SR, Nassu RT, Feijó GLD, Silva LOC, Torres RA, Siqueira F, Higa RH, Regitano LCA: **A genome scan for meat quality traits in Nelore beef cattle.** *Physiol Genomics* 2013, **45**(21):1012–1020.
65. Espigolan R, Baldi F, Boligon AA, Souza FR, Gordo DG, Tonussi RL, Cardoso DF, Oliveira HN, Tonhati H, Sargolzaei M, Schenkel FS, Carvalheiro R, Ferro JA, Albuquerque LG: **Study of whole genome linkage disequilibrium in Nelore cattle.** *BMC Genomics* 2013, **14**:305.

66. Bolormaa S, Pryce JE, Kemper KE, Hayes BJ, Zhang Y, Tier B, Barendse W, Reverter A, Goddard ME: **Detection of quantitative trait loci in *Bos indicus* and *Bos taurus* cattle using genome-wide association studies.** *Genet Sel Evol* 2013, **45**:43.
67. Bolormaa S, Hayes BJ, Hawken RJ, Zhang Y, Reverter A, Goddard ME: **Detection of chromosome segments of zebu and taurine origin and their effect on beef production and growth.** In *J Anim Sci*, Volume 89. United States: 2011 American Society of Animal Science; 2011:2050–2060.
68. Tizioto PC, Meirelles SL, Veneroni GB, Tullio RR, Rosa AN, Alencar MM, Medeiros SR, Siqueira F, Feijo GL, Silva LO, Torres RA Jr, Regitano LCA: **A SNP in *ASAP1* gene is associated with meat quality and production traits in Nelore breed.** In *Meat Sci*, Volume 92. England: 2012 Elsevier Ltd; 2012:855–857.
69. AOCS: **Rapid determination of oil/fat utilizing high temperature solvent extraction.** AOCS Official Proc. Am 5-04. In *Official methods and recommended practices of the AOCS*. 5th edition. Champaign, IL: American Oil Chemists' Society, AOCS; 2004.
70. Hara A, Radin NS: **Lipid extraction of tissues with a low-toxicity solvent.** *Anal Biochem* 1978, **90**(1):420–426.
71. Christie WW: **The composition and structure of milk lipids.** In *Developments in Dairy Chemistry*, Volume 2. Edited by Fox PF. New York, NY: Applied Science Publishers; 1983:1–35.
72. IUPAC: *Compendium of Chemical Terminology*, (the "Gold Book"). Compiled by McNaught A.D. and Wilkinson A. 2nd edition. Oxford, United Kingdom: Blackwell Scientific Publications; 1997.
73. Illumina Inc: **GenomeStudio Software v2011.1 Release Notes.** 2011 [http://supportres.illumina.com/documents/myillumina/1f2d00ae-b759-489b-82f9-0163ce085ae7/genomestudio_2011.1_release_notes.pdf]
74. Gilmour AR, Gogel BJ, Cullis BR, Thompson R: **ASReml User Guide Release 3.0.** In *Hemel Hempstead, HP1 1ES*. UK: VSN International Ltd; 2009.
75. VanRaden PM: **Efficient methods to compute genomic predictions.** *J Dairy* 2008, **91**(11):4414–4423.
76. Meuwissen TH, Hayes BJ, Goddard ME: **Prediction of total genetic value using genome-wide dense marker maps.** *Genetics* 2001, **157**(4):1819–1829.
77. Habier D, Fernando RL, Kizilkaya K, Garrick DJ: **Extension of the bayesian alphabet for genomic selection.** *BMC Bioinformatics* 2011, **12**:186.
78. Kizilkaya K, Fernando RL, Garrick DJ: **Genomic prediction of simulated multibreed and purebred performance using observed fifty thousand single nucleotide polymorphism genotypes.** *J Anim Sci* 2010, **88**(2):544–551.
79. Onteru SK, Fan B, Nikkila MT, Garrick DJ, Stalder KJ, Rothschild MF: **Whole-genome association analyses for lifetime reproductive traits in the pig.** *J Anim Sci* 2011, **89**(4):988–995.
80. Meyer LR, Zweig AS, Hinrichs AS, Karolchik D, Kuhn RM, Wong M, Sloan CA, Rosenbloom KR, Roe G, Rhead B, Raney BJ, Pohl A, Malladi VS, Li CH, Lee BT, Learned K, Kirkup V, Hsu F, Heitner S, Harte RA, Haeussler M, Guruvadoo L, Goldman M, Giardine BM, Fujita PA, Dreszer TR, Diekhans M, Cline MS, Clawson H, Barber GP, et al: **The UCSC Genome Browser database: extensions and updates 2013.** *Nucleic Acids Res* 2013, **41**(Database issue):D64–D69.
81. Flicek P, Ahmed I, Amode MR, Barrell D, Beal K, Brent S, Carvalho-Silva D, Clapham P, Coates G, Fairley S, Fitzgerald S, Gil L, García-Girón C, Gordon L, Hourlier T, Hunt S, Juettemann T, Kähäri AK, Keenan S, Komorowska M, Kulesha E, Longden I, Maurel T, McLaren WM, Muffato M, Nag R, Overduin B, Pignatelli M, Pritchard B, Pritchard E, et al: **Ensembl 2013.** *Nucleic Acids Res* 2013, **41**(Database issue):D48–D55.

doi:10.1186/1471-2156-15-39

Cite this article as: Cesar et al.: Genome-wide association study for intramuscular fat deposition and composition in Nelore cattle. *BMC Genetics* 2014 15:39.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

