# DATA NOTE



# Complete genome sequences of *Providencia* bacteriophages PibeRecoleta, Stilesk and PatoteraRojo



Steven Batinovic<sup>1\*</sup>, Hiu Tat Chan<sup>2,3</sup>, Jason Stiles<sup>3</sup> and Steve Petrovski<sup>2</sup>

# Abstract

**Objectives** *Providencia* is a genus of gram-negative bacteria within the order *Enterobacterales*, closely related to *Proteus* and *Morganella*. While ubiquitous in the environment, some species of *Providencia*, such as *P. rettgeri and P. stuartii*, are considered emerging nosocomial pathogens and have been implicated in urinary tract infection, gastrointestinal illness, and travelers' diarrhea. Given their intrinsic resistance to many commonly used antibiotics, this study aimed to isolate and sequence bacteriophages targeting a clinical *P. rettgeri* isolate.

**Data description** Here we report the complete genome sequence of three novel *Providencia* phages, PibeRecoleta, Stilesk and PatoteraRojo, which were isolated against a clinical *P. rettgeri* strain sourced from a patient in a metropolitan hospital in Victoria, Australia. The three phages contain dsDNA genomes between 60.7 and 60.9 kb in size and are predicted to encode between 72 and 73 proteins. These three new phages, which share high genomic similarity to two other *Providencia* phages previously isolated on *P. stuartii*, serve as important resources in our understanding about *Providencia* bacteriophages and the potential for future phage-based biotherapies.

Keywords Bacteriophage, Providencia, Genomics, Phage therapy

# Objective

*Providencia* are a genus of gram-negative bacteria of the family *Morganellaceae*, closely related to *Proteus* and *Morganella*. There are nine currently recognized species within the genus, with *P. rettgeri*, *P. stuartii* and *P. alcalifaciens* the most encountered in the context of human disease. They are commonly associated with nosocomial

\*Correspondence:

Steven Batinovic

batinovic-steven-yz@ynu.ac.jp

<sup>1</sup>Division of Materials Science and Chemical Engineering, Yokohama

National University, Yokohama, Kanagawa, Japan

<sup>2</sup>Department of Physiology, Anatomy, and Microbiology, La Trobe

University, Bundoora, VIC, Australia

<sup>&</sup>lt;sup>3</sup>Department of Microbiology, Royal Melbourne Hospital, Parkville, VIC, Australia



© The Author(s) 2023. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/. The Creative Commons Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

urinary tract infection [1], particularly in patients with long-term urinary catheters leading to "purple bag syndrome" [2], associated with wounds in burn patients [3], and linked to diarrhea and gastroenteritis in children and travelers [4, 5]. Members of *Providencia* are intrinsically resistant to commonly used antibiotics including colistin, the last resort antibiotic for multi-resistant gram-negative bacteria [6]. Consequently, *Providencia* spp., along with other pathogenic bacteria of the order *Enterobacterales* including *Klebsiella pneumoniae*, *Escherichia coli*, *Enterobacter* spp., *Serratia* spp., *Proteus* spp. and Morganella spp. are classified as Critical Priority 1 on the World Health Organizations global priority pathogens list requiring further research and development of new antibiotics [7].

Bacteriophages (phages), viruses that are capable of infecting and killing bacteria, represent a potential solution to this emerging problem. Bacteriophages are ubiquitous in both natural and artificial environments and predicted to be the most abundant biological entities on the planet [8]. Only 26 Providencia phage genomes have thus far been deposited in the NCBI GenBank as of July 2023. Here we have isolated three novel Providencia phages, named PibeRecoleta, Stilesk and PatoteraRojo, by screening worm farm effluent samples obtained from various locations in Victoria, Australia on a clinical isolate of P. rettgeri (strain 9744). Visible phage plaques (approximately 0.3 mm) were picked, subjected to a total of three rounds of purification to ensure each plaque resulted from a single virion, and propagated as previously described [9].

# **Data description**

DNA were extracted from 1 ml phage filtrates (>10<sup>10</sup> PFU ml<sup>-1</sup>) using a zinc chloride phenol:chloroformbased extraction [9, 10]. Isolated DNA (100 ng) were then prepared for sequencing using the NEBNext<sup>®</sup> Ultra<sup>™</sup> II DNA Library Prep Kit (NEB) followed by sequencing on an Illumina MiSeq using a v3 600-cycle kit (Illumina) to generate 300 bp paired-end reads (n=491,519-968,088 paired reads). Raw data were filtered using Trim Galore v0.6.4 with default settings (Q scores of  $\geq 20$ , with automatic adapter detection) [11], and assembled with SPAdes v3.9.0 with default settings [12]. The assembled genome of PibeRecoleta was 60,727 bp with a GC content of 49.3% (1248-fold read coverage), Stilesk was 60,924 bp with a GC content of 49.5% (2310-fold read coverage) and PatoteraRojo was 60,728 bp with a GC content of 49.4% (1410-fold read coverage).

Genome termini were identified to be 11-bp 5' cos overhangs (5'-GTGCGGAGAGC-3') on all three phages using PhageTerm v1.0.12 [13] and confirmed by manual inspection of raw reads [10]. Genes were identified using Glimmer3 [14] followed by manual adjustment. Genomes were annotated using a combination of searching against the NCBI Conserved Domain Database [15] and the Virfam Webserver [16]. No tRNA genes were detected using tRNAscan-SE v2.0 [17] or Aragorn v1.2.41 [18]. All software was used with default parameters. A total of 72–73 predicted coding sequences were identified in each of the phage genomes. Those of which could be assigned a function (~30%) were characteristically organized in functional modules involved in virion morphogenesis and lysis, and DNA replication and nucleotide metabolism (Data file 1, Data file 2) [19].

The three highly syntenic phages described here share high pairwise DNA sequence similarity (78.7-89.7%) as determined using VIRIDIC (Data file 3) [19, 20]. Examination of public databases for sequenced phage genomes similar to these three phages revealed two highly related Providencia phages, Redjac [21] and PSTCR9, which were both isolated on *P. stuartii*. (Data file 3) [19]. Redjac and PSTCR9 phages, which represent the only two members of the Redjacvirus genus, share over 70% intergenomic similarity to the three Providencia phages sequenced here, indicating PibeRecoleta, Stilesk and PatoteraRojo phages belong to the Redjacvirus genus (Data file 3). Members of the Redjacvirus are also known to share moderate nucleotide similarity (46-47%) and similar genomic organization to phages from the well-studied flagellotropic Chivirus genus (such as Enterobacteria phage Chi) which target members of the Enterobacteriaceae (Data file 3) [19, 20, 22]. Insights into the molecular mechanisms utilized by these Providencia phages may potentially be gained from work already performed on Chivirus phages.

# Limitations

*Providencia* phages PibeRecoleta, Stilesk and PatoteraRojo represent complete phage genomes. Our understanding of phages targeting *Providencia* is currently

Table 1 Overview of data files/data sets

Label	Name of data file/data set	File types (file extension)	Data repository and identifier (DOI or accession number)
Data file 1	Table S1 - Phage genome annotations	MS Excel file (.xlsx)	figshare (https://doi.org/10.6084/m9.figshare.23689797.v1) [19]
Data file 2	Figure S1 - Phage genome maps	PDF (.pdf)	figshare (https://doi.org/10.6084/m9.figshare.23689797.v1) [19]
Data file 3	Figure S2 - Similarity of <i>Providencia</i> phages to pub- licly available phages	PDF (.pdf)	figshare (https://doi.org/10.6084/m9.figshare.23689797.v1) [19]
Data set 1	Providencia phage vB-PreS-PibeRecoleta, complete genome	GenBank (.gbk)	NCBI nucleotide, https://identifiers.org/nucleotide:MT675124 [23]
Data set 2	Providencia phage vB-PreS-Stilesk, complete genome	GenBank (.gbk)	NCBI nucleotide, https://identifiers.org/nucleotide:MT675125 [24]
Data set 3	Providencia phage vB-PreS-PatoteraRojo, complete genome	GenBank (.gbk)	NCBI nucleotide, https://identifiers.org/nucleotide:MT675126 [25]
Data set 4	Raw reads of Providencia phages PibeRecoleta, Stilesk and PatoteraRojo	Raw sequence reads (.fastq)	NCBI BioProject, https://www.ncbi.nlm.nih.gov/bioproject?ter m=PRJNA1004027 [26]

## Acknowledgements

We thank the La Trobe University Genomics Platform for their 2200 TapeStation service.

## Author contributions

S.B, H.T.C and S.P conceived and designed the study. H.T.C and J.S isolated the phages. S.B purified the phage DNA, sequenced, and assembled the phage genomes. S.B wrote the manuscript. The author(s) read and approved the final manuscript.

#### Funding

S.B was funded by a JSPS Postdoctoral Fellowship (P20714).

#### Data Availability

The data described in this Data note can be freely and openly accessed on NCBI GenBank under the following accession numbers: *Providencia* phage v8-PreS-PibeRecoleta, MT675124; *Providencia* phage v8-PreS-Stilesk, MT675125; *Providencia* phage v8-PreS-PatoteraRojo, MT675126. Raw sequence reads are available in the associated BioProject PRJNA1004027. Associated Data files are available on figshare (https://doi.org/10.6084/ m9.figshare.23689797.v1). Please see Table 1 and references [19, 23–26] for details and links to the data.

# Declarations

#### **Competing interests**

S.B is the guest editor of the special issue. The remaining authors declare that they have no competing interests.

#### Abbreviations

No abbreviations.

**Ethics approval and consent to participate** Not applicable.

#### **Consent for publication**

Not applicable.

Received: 16 July 2023 / Accepted: 21 August 2023 Published online: 01 September 2023

#### References

- Cornaglia G, Frugoni S, Mazzariol A, Piacentini E, Berlusconi A, Fontana R. Activities of oral antibiotics on Providencia strains isolated from institutionalized elderly patients with urinary tract infections. Antimicrob Agents Chemother. 1995;39(12):2819–21.
- Dealler SF, Hawkey PM, Millar MR. Enzymatic degradation of urinary indoxyl sulfate by Providencia stuartii and Klebsiella pneumoniae causes the purple urine bag syndrome. J Clin Microbiol. 1988;26(10):2152–6.
- WENZEL RP, HUNTING KJ, OSTERMAN CA, SANDE MA. Providencia stuartii, a hospital pathogen: potential factors for its emergence and transmission. Am J Epidemiol. 1976;104(2):170–80.
- Yoh M, Matsuyama J, Ohnishi M, Takagi K, Miyagi H, Mori K, Park K-S, Ono T, Honda T. Importance of Providencia species as a major cause of travellers' diarrhoea. J Med Microbiol. 2005;54(11):1077–82.

- Gogry FA, Siddiqui MT, Sultan I, Haq QMR. Current update on intrinsic and acquired colistin resistance mechanisms in bacteria. Front Med. 2021;8:677720.
- Tacconelli E, Carrara E, Savoldi A, Harbarth S, Mendelson M, Monnet DL, Pulcini C, Kahlmeter G, Kluytmans J, Carmeli Y. Discovery, research, and development of new antibiotics: the WHO priority list of antibiotic-resistant bacteria and tuberculosis. Lancet Infect Dis. 2018;18(3):318–27.
- Batinovic S, Wassef F, Knowler SA, Rice DT, Stanton CR, Rose J, Tucci J, Nittami T, Vinh A, Drummond GR. Bacteriophages in Natural and Artificial environments. Pathogens. 2019;8(3):100.
- Chan HT, Ku H, Low YP, Batinovic S, Kabwe M, Petrovski S, Tucci J. Characterization of novel lytic bacteriophages of Achromobacter marplantensis isolated from a pneumonia patient. Viruses. 2020;12(10):1138.
- Batinovic S, Stanton CR, Rice DTF, Rowe B, Beer M, Petrovski S. Tyroviruses are a new group of temperate phages that infect Bacillus species in soil environments worldwide. BMC Genomics. 2022;23(1):777.
- 11. Martin M. Cutadapt removes adapter sequences from high-throughput sequencing reads. EMBnet J. 2011;17(1):10–2.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol. 2012;19(5):455–77.
- Garneau JR, Depardieu F, Fortier L-C, Bikard D, Monot M. PhageTerm: a tool for fast and accurate determination of phage termini and packaging mechanism using next-generation sequencing data. Sci Rep. 2017;7(1):1–10.
- Delcher AL, Bratke KA, Powers EC, Salzberg SL. Identifying bacterial genes and endosymbiont DNA with glimmer. Bioinformatics. 2007;23(6):673–9.
- Lu S, Wang J, Chitsaz F, Derbyshire MK, Geer RC, Gonzales NR, Gwadz M, Hurwitz DI, Marchler GH, Song JS. CDD/SPARCLE: the conserved domain database in 2020. Nucleic Acids Res. 2020;48(D1):D265–8.
- Lopes A, Tavares P, Petit M-A, Guérois R, Zinn-Justin S. Automated classification of tailed bacteriophages according to their neck organization. BMC Genomics. 2014;15(1):1–17.
- 17. Chan PP, Lowe TM. tRNAscan-SE: searching for tRNA genes in genomic sequences. In: *Gene prediction* Springer; 2019: 1–14.
- Laslett D, Canback B. ARAGORN, a program to detect tRNA genes and tmRNA genes in nucleotide sequences. Nucleic Acids Res. 2004;32(1):11–6.
- Batinovic S, Chan HT, Stiles J, Petrovski S. Data files 1–3 for BMC genomic Data Data note complete genome sequences of Providencia bacteriophages PibeRecoleta, Stilesk and PatoteraRojo. Figshare. 2023. https://doi. org/10.6084/m9.Figshare.23689797.
- 20. Moraru C, Varsani A, Kropinski AM. VIRIDIC—A novel tool to calculate the intergenomic similarities of prokaryote-infecting viruses. Viruses. 2020;12(11):1268.
- Onmus-Leone F, Hang J, Clifford RJ, Yang Y, Riley MC, Kuschner RA, Waterman PE, Lesho EP. Enhanced de novo assembly of high throughput pyrosequencing data using whole genome mapping. PLoS ONE. 2013;8(4):e61762.
- Gilchrist CL, Chooi Y-H. Clinker & clustermap. Js: automatic generation of gene cluster comparison figures. Bioinformatics. 2021;37(16):2473–5.
- Batinovic S, Chan HT, Stiles J, Petrovski S. Providencia phage vB\_PreS-PibeRecoleta, complete genome. 2020. NCBI Genbank. https://identifiers.org/ nucleotide:MT675124.
- Batinovic S, Chan HT, Stiles J, Petrovski S. Providencia phage vB\_PreS-Stilesk, complete genome. 2020. NCBI Genbank. https://identifiers.org/ nucleotide:MT675125.
- Batinovic S, Chan HT, Stiles J, Petrovski S. Providencia phage vB\_PreS-PatoteraRojo, complete genome. 2020. NCBI GenBank. https://identifiers.org/ nucleotide:MT675126.
- Batinovic S, Chan HT, Stiles J, Petrovski S. Raw reads of Providencia phages PibeRecoleta, Stilesk and PatoteraRojo. 2023. NCBI. https://www.ncbi.nlm.nih. gov/bioproject?term=PRJNA1004027.

## **Publisher's Note**

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.