

RESEARCH ARTICLE

Open Access



Population genetic structure and habitat connectivity for jaguar (*Panthera onca*) conservation in Central Belize

Angelica Menchaca^{1,2*} , Natalia A. Rossi^{2,3}, Jeremy Froidevaux¹, Isabela Dias-Freedman⁴, Anthony Caragiulo², Claudia Wultsch^{2,6}, Bart Harmsen^{5,7,9}, Rebecca Foster^{5,9}, J. Antonio de la Torre¹⁰, Rodrigo A. Medellin⁸, Salisa Rabinowitz² and George Amato^{2*}

Abstract

Background: Connectivity among jaguar (*Panthera onca*) populations will ensure natural gene flow and the long-term survival of the species throughout its range. Jaguar conservation efforts have focused primarily on connecting suitable habitat in a broad-scale. Accelerated habitat reduction, human-wildlife conflict, limited funding, and the complexity of jaguar behaviour have proven challenging to maintain connectivity between populations effectively. Here, we used non-invasive genetic sampling and individual-based conservation genetic analyses to assess genetic diversity and levels of genetic connectivity between individuals in the Cockscomb Basin Wildlife Sanctuary and the Maya Forest Corridor. We used expert knowledge and scientific literature to develop models of landscape permeability based on circuit theory with fine-scale landscape features as ecosystem types, distance to human settlements and roads to predict the most probable jaguar movement across central Belize.

Results: We used 12 highly polymorphic microsatellite loci to identify 50 individual jaguars. We detected high levels of genetic diversity across loci ($H_E = 0.61$, $H_O = 0.55$, and $N_A = 9.33$). Using Bayesian clustering and multivariate models to assess gene flow and genetic structure, we identified one single group of jaguars ($K = 1$). We identified critical areas for jaguar movement that fall outside the boundaries of current protected areas in central Belize. We detected two main areas of high landscape permeability in a stretch of approximately 18 km between Sittee River Forest Reserve and Manatee Forest Reserve that may increase functional connectivity and facilitate jaguar dispersal from and to Cockscomb Basin Wildlife Sanctuary. Our analysis provides important insights on fine-scale genetic and landscape connectivity of jaguars in central Belize, an area of conservation concern.

Conclusions: The results of our study demonstrate high levels of relatively recent gene flow for jaguars between two study sites in central Belize. Our landscape analysis detected corridors of expected jaguar movement between the Cockscomb Basin Wildlife Sanctuary and the Maya Forest Corridor. We highlight the importance of maintaining already established corridors and consolidating new areas that further promote jaguar movement across suitable habitat beyond the boundaries of currently protected areas. Continued conservation efforts within identified corridors will further maintain and increase genetic connectivity in central Belize.

Keywords: Conservation genetics, Population structure, Jaguar, Felidae, Landscape permeability, Functional habitat connectivity

* Correspondence: angelica.menchaca@bristol.ac.uk; gamato@amnh.org

¹School of Biological Sciences, the University of Bristol, Bristol, UK

²Sackler Institute for Comparative Genomics, American Museum of Natural History, New York City, USA

Full list of author information is available at the end of the article



Background

The jaguar (*Panthera onca*), listed as near threatened by the IUCN Red List of Threatened, species [1], is impacted by the depletion of wild prey, direct persecution and habitat loss [2]. As small populations become isolated by the fragmentation of habitat, jaguars become more vulnerable to demographic fluctuations, environmental perturbations, and inbreeding depression. Effective conservation planning for the jaguar requires comprehensive studies that effectively link the landscape with genetic structure and gene flow of the species at risk [3, 4]. Knowledge of the demographic history, movement patterns, behaviour and genetic diversity of a species are critical components to make adequate conservation and landscape planning decisions [5, 6].

Jaguars are elusive top predators with large individual home ranges and occurring at low densities in Neotropical forests [7]. After a century of persecution and suffering habitat loss, jaguar range decreased by 48% for the entire Americas and 85% outside Amazonia, [2, 8]. Furthermore, jaguar core areas contracted by 33% in the Gran Chaco [9]. To ensure population viability, the remaining fragmented patches of jaguar habitat need to be connected via a network of corridors that ensure demographic and genetic exchange [6, 10, 11]. Habitat deterioration, human-wildlife conflict and the complex dynamics of jaguar populations are rapidly shaping current patterns of genetic diversity [3, 12, 13].

The establishment of corridors to improve population connectivity, particularly in Mesoamerica, has been among some of the most important efforts to prevent the loss of biodiversity in the world's biologically richest regions [14]. One of the jaguar populations with the highest density in Mesoamerica can be found in the forests of Belize, a core and critical area for the species throughout its range [15]. However, jaguars in Belize are mainly threatened by overhunting of wild prey, direct persecution by humans, and habitat loss [16, 17].

According to the Belize National Protected Areas System Plan [18], 36% of Belize's land territory is under conservation management. In particular, with its 514 km² of protected forests, the Cockscomb Basin Wildlife Sanctuary harbours one of the largest concentrations of jaguars in the country [15, 19]. The Cockscomb Basin Wildlife Sanctuary is part of a larger conglomerate of protected areas, the Maya Mountains, forming the largest block of contiguous connected forest in Belize. This area is connected to the northern Selva Maya that extends over Belize to northern Guatemala and southeastern Mexico through a network of forest reserves, ending in an unprotected bottleneck stretch of forest in Central Belize.

The Maya Forest Corridor (formerly the Central Belize Corridor) was established in 2013 by the government of Belize and represents the only remaining source of

biological connectivity between the Selva Maya forests in northern Belize and the protected areas of the Maya Mountain Massif in southern Belize [16, 20]. This corridor extends over 750 km² and is comprised of mostly private lands but also communities, and protected areas including: the Labouring Creek Jaguar Corridor Wildlife Sanctuary (LCJCWS), the Peccary Hills National Park, and the Manatee Forest Reserve on national land and private protected areas such as Runaway Creek and Monkey Bay [20]. The Maya Forest Corridor is considered the most critical and important corridor of the Belize National Protected Areas Systems [20]. Within Central America, the Maya Forest Corridor has been considered the corridor with the highest probability of jaguar presence but also with the highest deforestation rate for large-scale agricultural developments, which have resulted in more than 65% reduction of natural habitat over the past decade, and as such in urgent need of protection [21, 22].

Range-wide movement corridors have been established as a major tool to improve population connectivity and thus aid the persistence of the jaguar across its range [23]. Conservation efforts have focused on a broad-scale and would benefit from information gained at a finer scale, especially across heterogeneous or fragmented landscapes [4, 11, 24]. The effective collaboration among scientists, practitioners, non-governmental organisations and politicians will tap the full potential of corridor projects and conservation actions across the jaguar's range.

Former non-invasive genetic studies on Neotropical felids in Belize confirmed the importance of Selva Maya jaguars and showed that jaguars sampled in Belize and northern Guatemala formed one genetic cluster with high levels of genetic diversity and gene flow between sites [13]. A countrywide non-invasive genetic study focusing on jaguars, pumas (*Puma concolor*) and ocelots (*Leopardus pardalis*) in Belize also detected moderate to high levels of genetic diversity and gene flow for jaguars but also indicated that fine-scale genetic subdivision was present between the most northern and southern sites within the country [25, 26]. The study suggested that human-altered landscapes adjacent to these sites most likely start to negatively impact jaguar movement and gene flow and further recommended continued genetic monitoring of these areas of conservation concern, including central Belize and the Maya Forest Corridor, which represents a critical link between jaguar habitats in northern and southern Belize.

Here, we present a fine-scale conservation genetic study assessing genetic structure and patterns of landscape connectivity for jaguars in central Belize to identify areas at risk. Using non-invasive genetic sampling and faecal DNA, we investigate population genetic structure, levels of genetic diversity, inbreeding and gene flow.

Additionally, we compared landscape features to examine landscape permeability for jaguars between the Cockscomb Basin Wildlife Sanctuary and the Maya Forest Corridor in central Belize.

Results

Genetic variation

A total of 536 scat samples collected across two areas in central Belize were positively matched to *P. onca*. (*n* = 414 Cockscomb Basin Wildlife Sanctuary; *n* = 126 Maya Forest Corridor). Other identified species included *Puma concolor*, *Leopardus wiedii*, *Leopardus pardalis*, *Herpailurus yagouaroundi*, and *Canis familiaris*. Genotyping revealed a total of 50 unique multilocus genotypes (37 from the Cockscomb Basin Wildlife Sanctuary and 13 from the Maya Forest Corridor); these included 41 prospective males, 3 prospective females and 6 unidentified genders (Additional file 3). MICROCHECKER detected three loci (FCA212, FCA229, and FCA075), showing signs of a null allele, but did not find evidence of scoring mistakes or large allele dropout (Additional file 1).

Twelve microsatellite loci were successfully amplified with a mean expected heterozygosity $H_E = 0.61$ ($SD = 0.042$), and a mean observed heterozygosity $H_O = 0.55$ ($SD = 0.05$). The mean number of alleles per locus ($N_A = 9.33$) ranged from 3 to 10 (Table 1). The mean polymorphic information content $PIC = 0.642$. We determined the geographical coordinates assigned to each individual by averaging the coordinates of all the samples corresponding to that particular individual. Tests for departure from Hardy-Weinberg Equilibrium followed by Bonferroni

correction were variable for each locus, with four loci deviating from HWE (Table 1). This deviation could be explained by a deficit of heterozygotes within the population potentially caused by inbreeding or by the presence of null alleles ($FIS = 0.22$, p -value = 0.001). Furthermore, this test showed evidence of low genetic differentiation ($FST = 0.021$, P -value = 0.007; $FIT = 0.237$, P -value = 0.001); linkage disequilibrium was not significant for any pair of loci. The DAPC of genetic diversity showed overlapping of the two sites, indicating overlapping of allele frequencies and little differentiation between groups (Fig. 1c). The AMOVA analysis revealed that less than 2% of genetic variance occurred among individuals in the Cockscomb Basin Wildlife Sanctuary and individuals in the Maya Forest Corridor; and showed low levels of genetic differentiation between groups ($FST = 0.015$, P -value = 0.026). Results from the Mantel test showed significant evidence of isolation by distance ($R_{xy} = 0.167$, p -value = 0.010; Fig. 1b).

Population structure and relatedness

Data analysis using STRUCTURE revealed that $K = 1$ had the highest mean probability of density value, and $K = 4$ had the highest delta- K value (Fig. 1a). This was consistent with the results from TESS, where $K = 1$ also had the highest probability ($\Delta K = 10.15$). In both cases, no clear pattern of genetic structure can be observed when rendering the assignment probability in bar plots (Fig. 1a). Results from GENELAND revealed that $K = 1$ also had the highest probability in 8 of 10 runs, and the final map does not show a clear population boundary between sampling sites (Fig. 1d). The DAPC analysis showed that the lowest BIC value (68.42) corresponded to $K = 2$ and is represented in a single discriminant function; however, the BIC difference between $K = 2$ and $K = 1$ is negligible (~ 1).

We analysed the performance of seven relatedness estimators to assess the degree of resolution expected in our dataset. Mean relatedness amongst individuals from the Maya Forest Corridor was -0.046 ± 0.068 ($SE = 0.008$) and from Cockscomb Basin Wildlife Sanctuary -0.15 ± 0.086 ($SE = 0.003$). Amongst all individuals mean relatedness was $-0.01 \pm SD 0.08$ ($SE = 0.002$). Overall, individuals from the Maya Forest Corridor were more closely related to each other than to those in the Cockscomb Basin Wildlife Sanctuary.

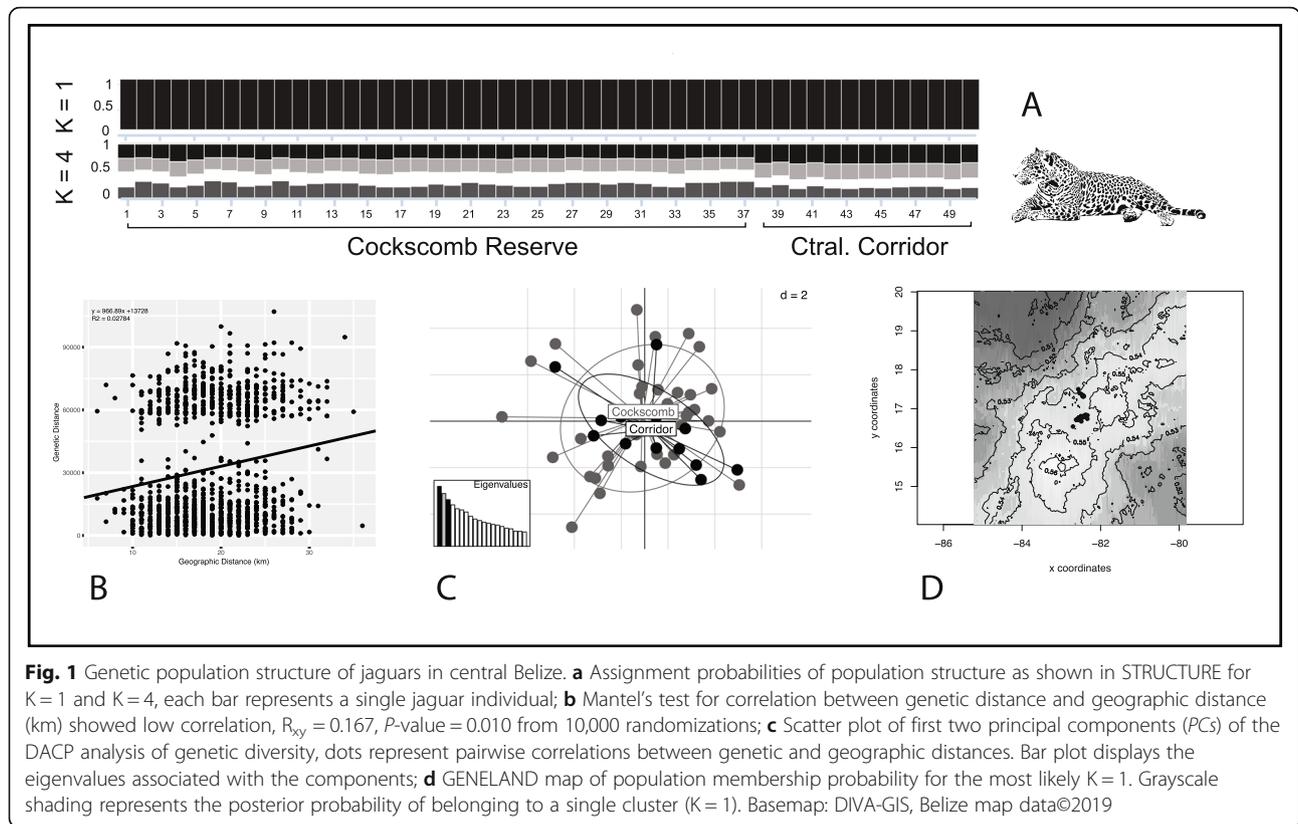
Landscape permeability

Paths of predicted jaguar movement were detected across the two study areas in central Belize (Fig. 2, Additional file 4). We identified important paths that have high potential to facilitate jaguar movement between the Cockscomb Basin Wildlife Sanctuary and the Maya Forest Corridor in central Belize. In particular, we detected two main areas potentially centralizing jaguar movement

Table 1 The number of alleles (N_A), allelic richness (A_R), observed (H_O) and expected heterozygosity (H_E), fixation index (F_{IS}), its standard error (SE), and P -value for the test of Hardy-Weinberg equilibrium for 12 microsatellite loci amplified for 50 jaguars

Locus	N_A	AR	H_O	H_E	F_{IS}	SE	P -value
FCA032	5	3.523	0.540	0.629	0.167	0.023	0.219
FCA100	5	3.687	0.520	0.597	0.147	0.056	0.085
FCA124	4	2.989	0.571	0.653	0.119	0.178	0.507
FCA126	6	4.454	0.766	0.709	-0.103	0.035	0.475
FCA212	2	1.989	0.163	0.273	0.437	0.355	0.015
FCA229	6	4.172	0.596	0.729	0.240	0.313	< 0.0001
FCA096	6	4.565	0.778	0.729	-0.051	0.139	0.584
FCA132	2	1.572	0.085	0.081	0.342	0.005	1.000
FCA275	3	2.995	0.532	0.643	0.217	0.111	0.010
FCA075	10	6.852	0.612	0.844	0.261	0.051	0.001
FCA208	7	5.533	0.833	0.767	-0.028	0.042	0.926
FCA225	4	3.691	0.524	0.585	0.238	0.024	0.144
	9.33		0.55	0.61	0.22		0.001

Bold values indicate loci not in Hardy-Weinberg equilibrium ($p < 0.01$) following Bonferroni correction. Estimates across all loci in italics



from Sibun and Sittee River Forest Reserves to Manatee Forest Reserve (– 88.5830 W, 17.0623 N; and – 88.4814 W, 17.0112 N), and vice versa, along a stretch of approximately 18 km adjacent to Hummingbird highway (Fig. 2). The areas connecting one reserve to the other ranges from approximately 25 m to 7 km.

Circuitscape analysis identified other critical paths of predicted jaguar movement that fall outside the boundaries of the Manatee Forest Reserve and the Maya Forest Corridor. In particular, Circuitscape detected high-current paths for jaguar movement in approximately 125 km² of unprotected habitat east of Manatee Forest Reserve and 70 km² of unprotected habitat west of the same reserve (Fig. 3, Additional file 4). Furthermore, Circuitscape found paths of high current density for jaguar movement both east and west of Sittee River Forest Reserve and northeast of Cockscomb Basin Wildlife Sanctuary and Mango Creek in an area of approximately 160 km² (Fig. 3).

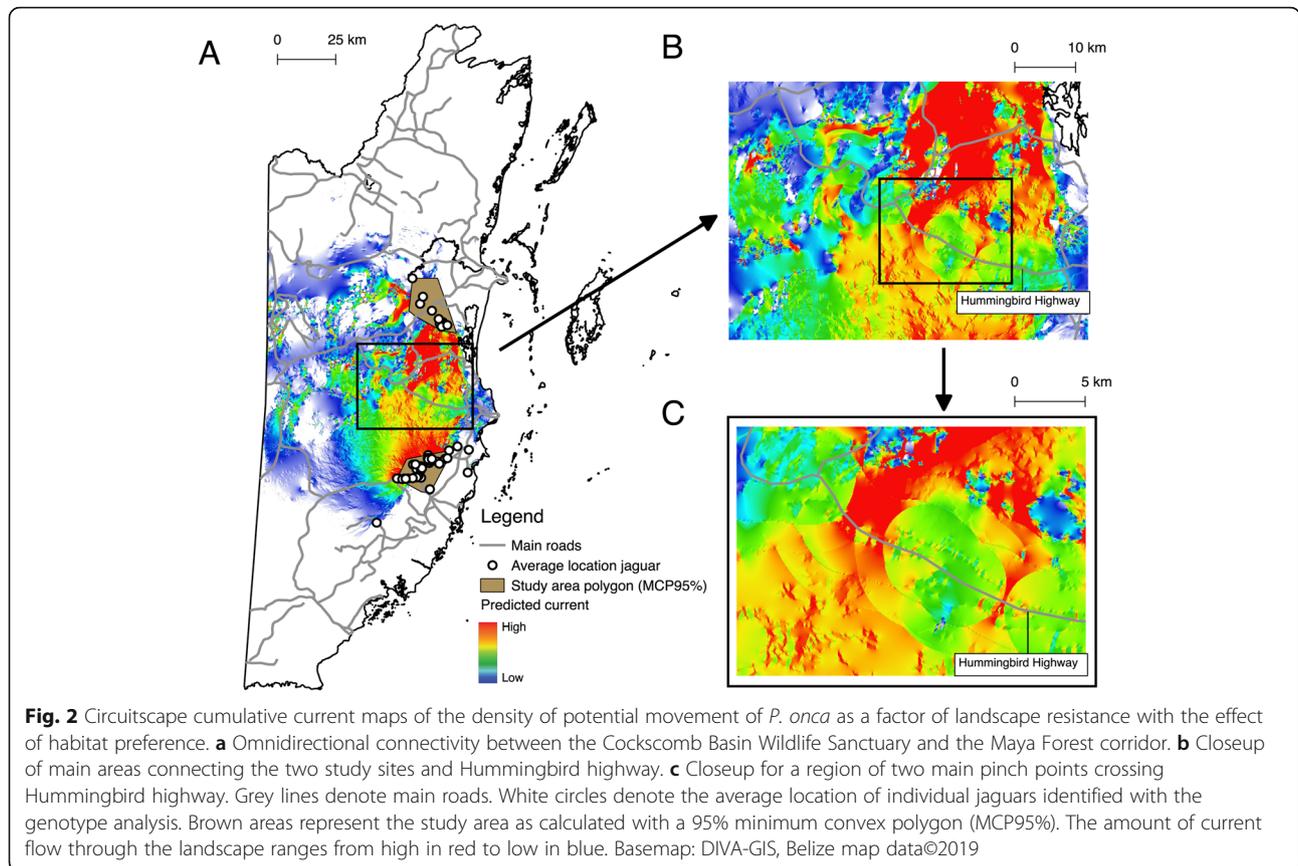
Discussion

Population genetics

This study assessed genetic diversity and structure for jaguars across two study sites in central Belize. Using twelve polymorphic microsatellite loci, we successfully identified 50 jaguar individuals corresponding to 41 males, 3 females and 6 individuals of undetermined

sexes. The relatively low number of females could be explained by the sampling methodology rather than reflecting the proportion of sexes in the area. Sampling was primarily conducted along trails, which tend to be dominated by males [27]; because females could be more elusive, have smaller home ranges, hide their scats and avoid crossing open spaces and wide paths [4, 5, 28–30] this method could favour the sampling of male scats and therefore bias the analysis towards the more frequently observed sex. Studies on dispersal in large felids show that males are the dispersing sex, while females tend to be more philopatric [11, 13, 26, 31, 32]; other measurements for genetic differentiation between sexes and more female scat samples are necessary to confirm sex-biased dispersal in this area.

We estimated high levels of genetic diversity and gene flow from data derived from microsatellite analysis and did not detect genetic population structure between the Cockscomb Basin Wildlife Sanctuary and the Maya Forest Corridor (Fig. 1). Overall, our results show evidence of high gene flow between sites with low population differentiation and a lack of heterozygosity within certain loci (Table 1). The results from the Bayesian clustering analyses assigned all 50 jaguar individuals to one single genetic cluster and did not show any clear patterns of population structure (Fig. 1a). Our results are consistent with former studies that revealed jaguar movement

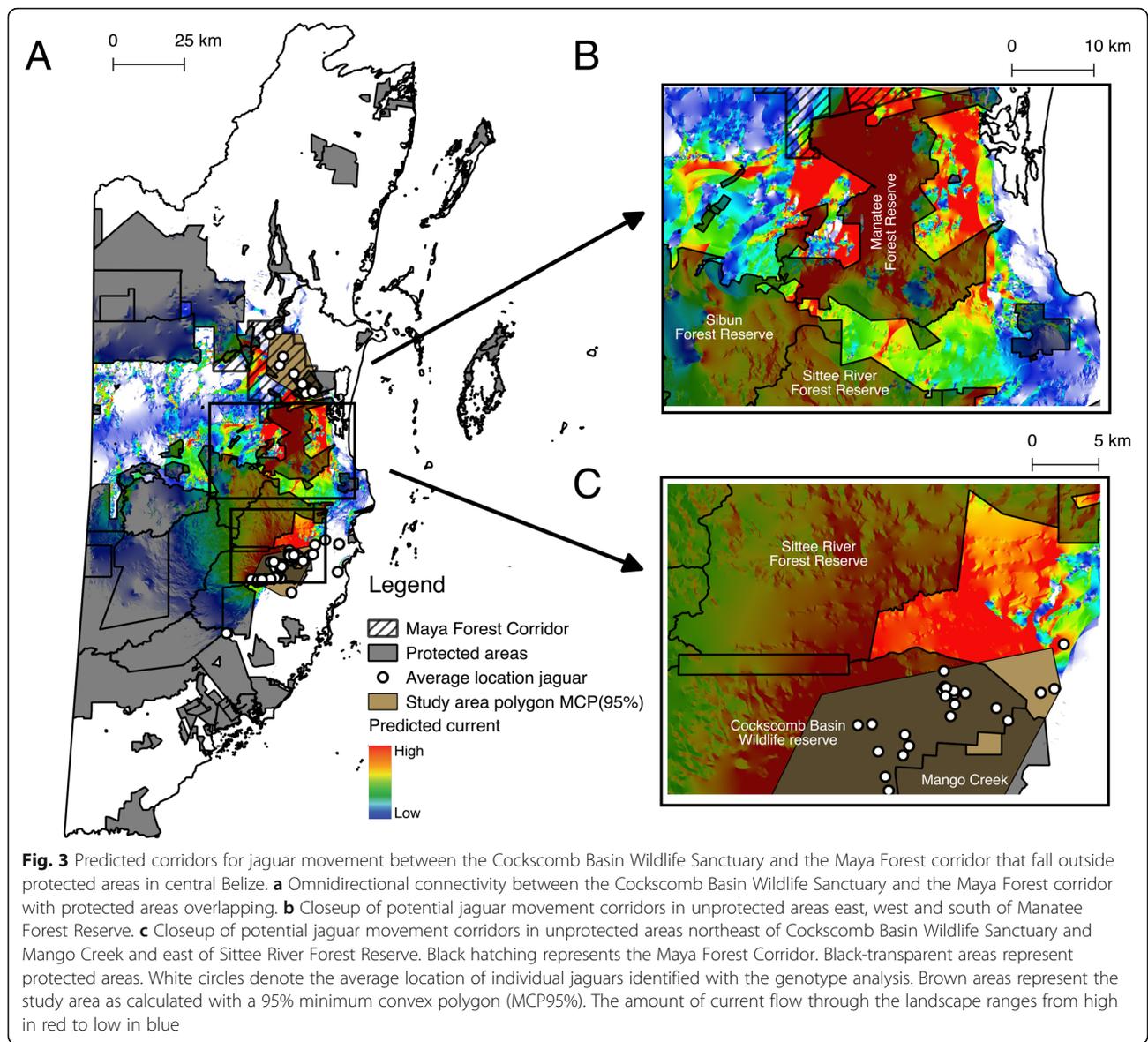


across the landscape in Belize and relatively high gene flow [26]. However, the spatial scale of this study and biases towards the sampling of jaguar males could potentially limit some of our results. A small spatial scale relative to the movement of sampled jaguars (mostly males) could make it less likely to detect subtle patterns of genetic population structure. When expanding the geographic scope to the entire country, previous studies revealed fine-scale genetic subdivision of jaguar populations between the most northern and southern sites within the country [25, 26].

In addition, the power of Bayesian clustering to detect population structure has been shown to decrease in accuracy at very low levels of population differentiation [33] as the number of estimated populations can be affected by a violation of model assumptions and cryptic relatedness [34]. Bayesian clustering methods rely on genetic information to ascertain population membership and operate by minimising Hardy-Weinberg and linkage disequilibria [33]. Additionally, properties of our data as sample size, number of loci, polymorphisms, and null alleles could have also influenced their performance [35, 36]. Support for a single population was achieved with GENELAND and DAPC analyses, which indicated a single cluster as the most probable number of populations

(Fig. 1c, d). GENELAND assigned individuals to each population considering the sampling locations and measurements of genetic differentiation; as this method considered spatial autocorrelation and is more able to detect low levels of genetic differentiation, it more accurately reflects the true K [37]. The closer relatedness of jaguars from the Maya Forest Corridor than that found for jaguars in the Cockscomb Basin Wildlife Sanctuary could potentially indicate more stable spatial groups or barriers to movement at the Maya Forest Corridor compared to the Cockscomb Basin Wildlife Sanctuary.

The seven-year sampling gap between the collection of scats from the two localities (Cockscomb Basin Wildlife Sanctuary in 2003, and Maya Forest Corridor in 2010) and the sampling bias towards the more dispersing sex (males) could have impacted the results of this study. For instance, our genetic results could reflect dispersal of young males into new territories, hindering possible patterns of subtle genetic structuring among females. Limited or sex-biased dispersal between sampling localities could further contribute to genetic differentiation between the localities. Studies conducted with radio telemetry show that jaguars depend on large patches of habitat and can have home ranges that surpass 100 km² [4, 7, 15, 16, 38] however, females have smaller home



ranges and tend to avoid roads and human-dominated landscapes at a higher degree, showing preference for intact forests [4, 11, 39]. Although having large home ranges and the ability to move considerable distances, jaguars tend to avoid human-dominated areas and show gender-specific differences [4, 27, 39]. Genetic subdivision due to limited gene flow between jaguar populations has been suggested to be primarily caused by habitat fragmentation, loss and human disturbances [13]. Other studies have also demonstrated that jaguars are highly sensitive to habitat fragmentation in human-dominated landscapes and that although dispersal capabilities of the species may slow the effect of drift, the effect of large-scale habitat loss and fragmentation may contribute to genetic differentiation within a short period of time [3, 6].

Landscape connectivity

The relatively high levels of gene flow and low genetic differentiation found in our study attest to the successful connectivity between jaguars in the Maya Forest Corridor and the Cockscomb Basin Wildlife Sanctuary, which were a continuum of jaguar habitat in the distant past but that still maintain a single genetic population. Movement corridors are essential to maintain the genetic connectivity for wide-ranging carnivores like jaguars [23, 40] and are an example of an effective conservation strategy that could be implemented in other areas of the species range, such as those in the Atlantic Forest of South America, where there is a lack of genetic and landscape connectivity among isolated remnant jaguar populations [3, 24, 41, 42]. The effect of jaguar corridors on the species conservation is an effective conservation strategy

that has also contributed significantly to the conservation and connectivity of a larger number of co-occurring species [40].

Anthropogenic barriers and accelerated rates of habitat conversion into agricultural lands are likely to alter gene flow between core jaguar areas if these are not effectively connected [3, 6, 42, 43]. In this study, we identified several areas in central Belize that are likely to be used by jaguars as they move through the landscape and that represent potential locations for priority management to ensure continued gene flow (Figs. 1, 2). In particular, the negative impact of roads on jaguar populations should be considered to improve existing corridors or in the design of new ones; for example, roads can block the movement of wildlife, and also open up areas to hunters [4, 41, 43].

Using circuit theory, we incorporated landscape features to predict suitable pathways for jaguar movement between the Maya Forest Corridor and the Cockscomb Basin Wildlife Reserve. We pinpointed two main areas of connectivity between Sittee River Forest Reserve and Manatee Forest Reserve along 'Hummingbird Highway', a clear anthropogenic boundary between the two sites (Fig. 2). These critical areas are important to maintain connectivity outside the boundaries of Manatee, Sibun and Sittee River Forest Reserves. These results are preliminary and limited by the geographic sampling of our study, in addition to the scarcity of information regarding fine-scale habitat use of jaguars in the area. Furthermore, even though widely utilized, expert knowledge approaches alone are limited and need to incorporate empirical parameterization of the permeability surface to achieve greater accuracy [44–46]. Studies looking at jaguar occurrence, density, behaviour and movement patterns in the region will be critical to creating more accurate connectivity models and improving the reliability of landscape analyses. Studies in central Belize with camera trapping and tracking with GPS radio-collared individuals will confirm the frequency of use of such areas as a passage between reserves and would provide more information on the impact of Hummingbird highway on jaguar movement. The standard means of studying jaguar populations through the deployment of camera traps in Belize [19, 29] can also be augmented with technologies such as three-dimensional remote sensing, which critically improves functional connectivity estimates compared to conventional land cover data [47].

Conclusion

Our results provide a screenshot of genetic patterns of animals whose scats were sampled during 2003–2011. Our study found high levels of gene flow and the presence of a single jaguar population inhabiting the Maya Forest Corridor and the Cockscomb Basin

Wildlife Sanctuary. We identified important areas that have great potential to facilitate jaguar movement and maintain connectivity between the Maya Forest Corridor and the Cockscomb Basin Wildlife Reserve in central Belize (Fig. 2). Our landscape connectivity analysis provides critical information useful to mitigate future effects of habitat fragmentation and anthropogenic activities on jaguar movement and gene flow. The results of this research provide a basis for conservation decisions on jaguars, having to travel through human-dominated landscapes. Non-invasive genetic monitoring has a high potential to provide critical information for conservation planning for threatened wildlife in Belize, as shown in this study. Future studies with large genome-wide datasets will be useful to evaluate fine-scale population structure and will allow investigating loci exhibiting ecologically relevant adaptation [48]. However, in view of the urgent need of understanding genetic variability and population structure, and the relatively ease and reduced cost of using a few loci to understand patterns of genetic diversity, we recommend using the suit of microsatellite markers used in this study (see Additional file 2) to compare jaguar populations in different countries. Furthermore, we recommend sampling in western and southern Belize to better predict habitat connectivity across the Maya Mountain Massif and between this area and the Maya forests in the north. By combining analyses on population genetics, landscape ecology and three-dimensional remote sensing, we will achieve a better understanding of the influence of landscape features and anthropogenic activities on jaguar movement and gene flow across Belize.

The difficulty of studying jaguars in the wild has limited researchers in their efforts to resolve conservation and management issues. Furthermore, given the new environmental challenges and other anthropogenic threats that jaguars and other wildlife face, it is important to support the Belize government to maintain and consolidate corridors that connect suitable habitat and allow the movement of top predators like jaguars. The efforts that international and national conservationists, scientists, local communities and politicians have placed in the last 15 years have helped maintain a relatively healthy jaguar population in central Belize. We encourage continued genetic and ecological monitoring in coordination with management actions taking place in Mexico, Guatemala and other Central American countries to promote connectivity among isolated remnant jaguar populations. Consolidating jaguar corridors and expanding natural protected areas is fundamental to make it a long-term success and one that requires local and international support.

Methods

Sampling

Scat collection was conducted by Panthera Belize in two sites (Fig. 2): 1) the Cockscomb Basin Wildlife Sanctuary and some areas that fall beyond the boundaries of this natural reserve, between 2003 and 2007; and 2) the Maya Forest Corridor between 2009 and 2011. Sampling was conducted as part of the Global Felid Genetics Program. The Maya Forest Corridor (17.349140° N, 88.455310° W, 50 m elevation) has been identified as an important link between jaguar populations in northern and southern Belize [20]. The Cockscomb Basin Wildlife Sanctuary (16.7162° N, 88.6608° W, 500 m elevation) supports the highest density of jaguars in Mesoamerica [15, 19]. These two areas play a crucial role in the maintenance of the Mesoamerican Biological Corridor, comprised of a network of protected areas stretching from Mexico to Panama [23]. A total of 852 scats were collected opportunistically along forest trails and unpaved roads. Samples were georeferenced at the time of collection and stored individually at room temperature using silica gel beads until DNA extractions. All faecal material has been deposited in the Sackler Institute for Comparative Genomics at the American Museum of Natural History, NYC.

DNA extraction and species identification

About 200 mg of the dry sample was shaved from each scat and used to extract genomic DNA using a QIAmp DNA extraction Stool Mini Kit (Qiagen Inc., Valencia, California, USA). Samples were screened to identify species via PCR using three mitochondrial gene regions including cytochrome *b* (H15149) [49, 50], *12S* rDNA (L1085 and H1259) [51] *16S* rDNA (L2513 and H2714) [51] and *16S_{cp}* (16S cp-F 16S cp-R) [51]. PCR amplifications were done using G&E Ready-to-go PCR Beads (GE Healthcare, Piscataway, New Jersey, USA) in 25ul reaction volumes containing 0.2uM of each primer, 0.3uL BSA and 1ul of DNA. PCR profiles for each reaction are available as supplementary material (see Additional file 2). All extractions and amplifications included negative controls. Cycle-sequencing was performed with the BidDye® Terminator v. 1.1 cycle Sequencing kit (Applied Biosystems, Lennik, Belgium) using the PCR primers. Amplified products were visualised and scored in an ABI 3730xl DNA Analyzer (Applied Biosystems, Carlsbad, CA). Sequences were aligned and manually corrected using GENEIOUS v.6.5 (Biomatters Ltd., Auckland, New Zealand). Species identification was confirmed by comparing consensus sequences to known felid references and by constructing a phylogenetic tree with 93% similarity using the Jukes-Cantor neighbourhood-

joining model [52]. Species ID was validated if at least three gene fragments successfully identified the same species.

Individual and sex identification

Samples identified as *P. onca* were genotyped for twelve polymorphic microsatellite loci (FCA 32, 75, 96, 126, 100, 124, 132, 208, 212, 229, 275, and 225) [53]. Each sample was screened 4 times to have enough replicates to assign the alleles correctly and to reduce artefacts caused by allele dropout, very difficult samples were screened up to 6 times. The genotypes were validated if they successfully scored at least three times. Samples that failed to amplify at least six loci were not considered in our analysis. We amplified sets of 2–3 primers in multiplex PCR reactions and FCA225 as singleplex (see Additional file 2). Amplification was performed using the Qiagen Multiplex Master Mix® (Qiagen Inc., Valencia, CA), with a final volume of 20ul following manufacturers recommendations.

PCR reaction conditions were optimised in the annealing temperature for each set of primers as follows: 95 °C for 15 min, followed by 13 cycles of 94 °C for 30s, 57.4–62.4 °C for 90s, 72 °C for 60s, followed by 32 cycles of 94 °C for 30s, 55–60 °C for 90 s, 72 °C for 60s and 60 °C for 30 min (see Additional file 2). Amplified products were visualised and scored in an ABI 3730xl DNA Analyzer (Applied Biosystems, Carlsbad, CA). PCR products were analysed using GeneMapper v5.0 following the quality index measure to validate genotypes (Miquel et al. 2006). In order to further validate the allele calling, we used FRAGMAN v.1.0.8 package in R [54]. Identification of unique multilocus genotypes was achieved using the ALLELEMATCH package in R v.2.5 [55], which accommodates for genotyping error and missing data by making pairwise comparisons and finding similarity scores for each pair of profiles and clustering similar ones. To determine if genotypes were not different individuals matched by chance, the cumulative probability of identity for unrelated individuals (PID) and siblings (Psib) was calculated. Genotypes with Psib < 0.010 were positively identified as single individuals [25, 56].

Gender was determined following the PCR-CTPP method for sex identification developed by Wei et al. (2008) based on zinc finger alleles (ZFX/ZFY). The two forward primers (ZF-1F and ZFY-2F) were fluorescently labelled with FAM and MAX, respectively. Amplification was performed using G&E Ready-to-go PCR Beads (GE Healthcare, Piscataway, New Jersey, USA) in a final volume of 25ul containing 2 mM of each primer and 5ul of DNA. The PCR conditions were as follows: 95 °C for 5 min, followed by 13 cycles of 95 °C for 45 s, 63 °C for 60s, 72 °C for 60s, followed

by 35 cycles of 95 °C for 45 s, 53 °C for 45 s, 72 °C for 60s and 72 °C for 10 min (see Additional file 2). DNA amplifications were visualised in a 5% agarose gel to confirm the molecular sexing. Amplified products were visualised and scored in an ABI 3730xl DNA Analyzer (Applied Biosystems, Carlsbad, CA). PCR products were analysed using GeneMapper v5.0.

Conservation genetics analyses

MICROCHECKER v 2.2.3 [57] was used to screen null alleles at each locus. Measures of genetic diversity as average number of alleles per locus (A), allelic richness (AR), observed (H_o) and expected heterozygosity (H_e) deviations from Hardy-Weinberg equilibrium (HWE), inbreeding coefficient (FIS), and multivariate analyses were performed using the ADEGENET package in R v.2.0.1 [58]. The rarefaction procedure implemented the HIERFSTAT package in R v 2.9.3.250 [59] was used to estimate the expected number of alleles (N_A) and to compare allelic richness (A_R). Genetic differentiation among sampled individuals was summarised in a Discriminant analysis of principal components (DAPC) based on the allele frequencies. Linkage disequilibrium (LD) between pairs of loci was performed in GENEPOP v. 4.2 [60] with default settings. We used GenAlix v6.0 [61] to test the occurrence of a positive correlation ($R_{xy} > 0$) between the genetic PHI_{PT} matrix and geographic distance so-called Isolation by Distance (IBD) via Mantel tests of matrix correspondence; we assessed the partitioning of genetic variation between sampling localities with Analysis of Molecular Variance (AMOVA).

Population structure

We estimated population genetic structure using Bayesian assignment methods with STRUCTURE v2.3.4 [62], which assigns individuals to a number K of genetically homogeneous groups, based on the Bayesian clustering in accordance to the expected Hardy-Weinberg equilibrium and absence of linkage disequilibrium between loci. We ran STRUCTURE using the admixture model with correlated allele frequencies and sampling locations as spatial prior (LOCPRIOR) to allow sampling location to assist in the clustering, and we performed 20 independent runs for $K = 1-10$. We set a burn-in period of 100,000 and 1,000,000 MCMC iterations and assumed an admixture model with correlated allele frequencies. To determine the optimal number of clusters and render bar plots, we implemented the Evanno method [35] using POPHELPER package in R v1.2.1 [63]. Furthermore, we inferred spatial genetic structure with TESS v2.3.1 [64]. This program assumes that population memberships follow a hidden Markov random field model where the log-probability of an individual belonging to a

particular population, given the population membership of its closest neighbours, is equal to the number of neighbours belonging to this population [65]. We tested the CAR, and BYM models with linear trend surface to define the spatial prior for admixture [36]; we set a burn-in period of 100,000 and 1000,000 sweeps through 10 independent runs testing the maximal number of clusters from 1 to 10. To decide the optimal K , we plotted the deviance information criterion (DIC) against K_{max} . We also used a spatial clustering approach in GENELAND v4.0.6 [65] as an additional method to infer the number of populations and the spatial location of genetic discontinuities. This program allows using georeferenced individual multilocus genotypes to infer the number of populations and uses the spatial location of genetic discontinuities between those populations. We determined K across 20 independent runs with 1,000,000 MCMC iterations. Thinning was set at 100, allowing K to vary from 1 to 10. We used the correlated allele model and set the maximum rate of the Poisson process at 50 (the number of individuals), the maximum number of nuclei in the Poisson-Voronoi tessellation at 150 (three times the number of individuals), and the uncertainty of spatial coordinates of the collection at 25 m. We re-ran the analysis ten times to check for consistency across runs.

To further explore genetic diversity and structure among individuals, we reduced the dimensions via a Discriminant Principal Component Analysis (DAPC) without a priori group assignment using the ADEGENET package in R v2.0.1 [58, 66]. The tools implemented in DAPC allow solving complex population structures by summarising the genetic differentiation between groups while overlooking within-group variation, therefore achieving the best discrimination of individuals into pre-defined groups [67]. This multivariate method is useful to identify clusters of genetically related individuals when group priors are lacking. Estimation of clusters was performed by comparing the different clustering solutions using the Bayesian Information Criterion (BIC). We compared the results from the three Bayesian approaches and the DAPC to provide confidence in the spatial designation of genetic groupings.

Relatedness

Levels of genetic relatedness were calculated using seven estimators as implemented in the RELATED package in R v1.0 [68]. Pairwise relatedness was calculated using the estimators described by Queller and Goodnight, 1989 [80]; Li et al., 1993 [81]; Ritland, 1996 [82]; Lynch and Ritland, 1999 [83] and Wang, 2002 [84], as well as the dyadic likelihood estimator described in Milligan, 2003 [85] and the triadic likelihood estimator from

Wang, 2007 [72]. Genotyping errors and inbreeding estimations were incorporated into the model, and confidence intervals (95%) were obtained through bootstrapping across loci. Allele frequencies were used to simulate pairs of individuals of known relatedness based on Parent-Offspring, Full siblings, Half siblings and Unrelated individuals.

Landscape connectivity

We used ArcGIS Desktop v10 to model resistance surfaces of the landscape to jaguar movement and CIRCUITSCAPE v3.5 [69] to identify the most probable routes for dispersal and gene flow between the Cockscomb Basin Wildlife Sanctuary and the Maya Forest Corridor by. Our analysis describes the connectivity and permeability of the landscape between the Cockscomb Basin Wildlife Sanctuary and the Maya Forest and was used to predict all possible paths connecting these two areas. We created a resistance raster surface at a 50 m resolution by combining data on ecosystem type, human settlement and roads. Spatial data were obtained from the Biodiversity and Environmental Resource Data System of Belize (BERDS) and included ecosystem types (2017), protected areas (2015), roads (2013), and human settlements (2014) [70].

The probability of jaguar occurrence has been positively associated with forest cover and negatively associated with human activities [71]. We assumed that landscape features vary in their importance to promote connectivity for jaguars; for example, lowland broad-leaved forests may promote jaguar movement while human settlement may limit it. To represent the variation in importance for connecting different areas, we created a surface of cost values of habitat preference (Table 2) based on published scientific literature and expert knowledge of nine jaguar scientists with experience working in Belize and South-Eastern Mexico. Although basing cost values on expert knowledge as quantitative information is controversial, it has been commonly used as a surrogate when empirical data is limited or unavailable [8, 69–72, 86–89]. To gather the information, we asked jaguar scientists to rank habitat preference according to their empirical and theoretical knowledge on habitat preference and resistance features to allow movement of jaguars across the landscape. The expert’s knowledge was supplemented with empirical data of jaguar habitat use [4, 16, 23, 39, 73–79]. Therefore, by expert knowledge, we mean the experts’ personal judgement that agrees with empirical data.

Each pixel was assigned a cost value in a scale of 1–9, which represents the relative effort required to move from one point to another (higher values

Table 2 Cost-values of jaguar habitat preference based on expert knowledge. Ecosystem names follow UNESCO’s classification system. Values range from 1 (not preferred) to 9 (highly preferred)

Landscape feature	Preference
Lowland broad-leaved wet forest	9
Submontane broad-leaved wet forest	8.8
Lowland broad-leaved moist forest	8.7
Submontane broad-leaved moist forest	8.7
Lowland broad-leaved moist scrub forest	7.4
Lowland broad-leaved wet forest: Steep	7.1
Submontane broad-leaved wet forest: Steep	7
Lowland broad-leaved dry forest	7
Submontane broad-leaved moist forest: Steep	6.9
Lowland broad-leaved moist forest: Steep	6.9
Lowland broad-leaved moist scrub forest: Steep	6.1
Mangrove and littoral forest	5.8
Lowland pine forest	5.7
Wetland	5.4
Submontane pine forest	5.4
Water	5.3
Shrubland	5.1
Lowland savanna	4.4
Submontane pine forest: Steep	3.9
<i>Anthropogenic feature</i>	
> 5 km distance from Highways	9
> 5 km distance from human settlements	9
> 3 and ≤ 5 km distance from Highways	6.5
> 3 and ≤ 5 km distance from human settlements	6.5
≤ 3 km distance from Highways	6.1
≤ 3 km distance from human settlements	5.4
Highway	3.3
Agricultural uses	2.7
Aquaculture	2
Wasteland	1.8
Urban	1

facilitate movement) (Table 2). The average locations of the identified jaguars were grouped in single polygons per study area and were calculated as the 95% minimum convex polygon (MCP). The resulting connectivity between polygons is represented as a cumulative current flow map where areas with higher predicted movement are coloured in red and areas of lower predicted movement in blue. The cumulative current maps represent all potential routes that an animal moving from one polygon could use to move across central Belize and potentially disperse to and from the Cockscomb Basin Wildlife Sanctuary to the Maya Forest Corridor (Fig. 2).

Supplementary information

Supplementary information accompanies this paper at <https://doi.org/10.1186/s12863-019-0801-5>.

Additional file 1: Figure S1. Probability graphs of K as calculated by Evanno et al. (2005). **Figure S2.** Box plots, and density plot of relatedness values for simulated pairs of individuals of known relatedness. **Figure S3.** Map of ecosystem types in Belize (2017). **Table S2.** Results from MICROCHECKER for 12 microsatellite loci.

Additional file 2: Table S1. Excel template with PCR profiles for species identification, microsatellite multiplexes and sex identification.

Additional file 3: Table S3. List of genotypes for 50 individual jaguars identified in the Cockscomb Basin Wildlife Sanctuary and the Maya Forest Corridor.

Additional file 4. TIF file to be open in GIS with cumulative current map projections of potential movement of *P. onca* between the Cockscomb Basin Wildlife Sanctuary and the Maya Forest Corridor, Belize.

Abbreviations

AMOVA: Analysis of Molecular Variance; BERDS: Biodiversity and Environmental Resource Data System of Belize; BIC: Bayesian information criterion; DAPC: Discriminant analysis of principal components; DNA: Deoxyribonucleic acid; IBD: Isolation by Distance; LCJWS: Labouring Creek Jaguar Corridor Wildlife Sanctuary; LD: Linkage disequilibrium; MCP: Minimum convex polygon; *P. onca*: *Panthera onca*; PC: Principal component; PCR: Polymerase chain reaction; rDNA: Recombinant DNA

Acknowledgements

We would like to thank the following jaguar experts for their input on habitat preferences used in this analysis: Gerardo Ceballos, Marcelo Aranda, Arturo Hernandez-Mijangos, Dulce M. Najera. We acknowledge Allison Devlin and Omar A. Figueroa for their contributions to the scat sample collection. We thank Orly Razgour, Angelo Soto-Centeno and Armando E. Quito for their helpful suggestions and support in the development of this study. We thank the laboratory technicians at AMNH and the field assistants for their hard work. We thank the reviewers for critical reading and thoughtful comments on this manuscript.

Authors' contributions

AM and NR analysed, interpreted the genetic and landscape data and wrote the original manuscript. JF performed the GIS simulations. IDF and AC conducted the species identification and part of the laboratory analyses. BH and RF conducted the sample collection and together with CW, ADT, RM provided critical feedback on the content of the manuscript. SR and GA supervised the project. All authors read and approved the final manuscript.

Funding

This work was conducted with the financial support of Panthera and the Sackler Institute for Comparative Genomics at the American Museum of Natural History. Financial supporters of the fieldwork include the Wildlife Conservation Society, the UK Natural Environmental Research Council, Panthera, the Liz Claiborne Art Ortenberg Foundation, the North England Zoological Society, Brevard Zoo, Woodland Park Zoo, the UK Darwin Initiative. AM acknowledges the scholarship provided by CONACYT (CVU 386490) through the program Maestrías en el Extranjero.

Availability of data and materials

All data generated or analysed during this study are included in this published article [and its supplementary information files].

Ethics approval and consent to participate

This research does not involve human subjects. Scat samples were obtained non-invasively in accordance with collection permits from the Belize Forest Department.

Consent for publication

This manuscript does not relate to an individual person.

Competing interests

The authors declare that they have no competing interests.

Author details

¹School of Biological Sciences, the University of Bristol, Bristol, UK. ²Sackler Institute for Comparative Genomics, American Museum of Natural History, New York City, USA. ³The Wildlife Conservation Society, New York City, USA. ⁴Columbia University in the City of New York, New York City, USA. ⁵Panthera, New York City, USA. ⁶City University of New York, New York City, USA. ⁷Environmental Research Institute, University of Belize, Belmopan, Belize. ⁸Instituto de Ecología, Universidad Nacional Autónoma de México, México City, México. ⁹Southampton University, Southampton, UK. ¹⁰School of Environmental and Geographical Sciences, University of Nottingham Malaysia, Semenyih, Malaysia.

Received: 17 August 2019 Accepted: 15 December 2019

Published online: 27 December 2019

References

- Quigley H, Foster R, Petracca L, Payan E, Salom R, Harmsen B. *Panthera onca* (errata version published in 2018). IUCN Red List Threat Species [Internet]. 2017 [cited 2019 Oct 21];e.T15953A1. Available from: <https://doi.org/10.2305/IUCN.UK.2017-3.RLTS.T15953A50658693.en>.
- Antonio De La Torre J, González-Maya JF, Zarza H, Ceballos G, Medellín RA. The jaguar's spots are darker than they appear: assessing the global conservation status of the jaguar *Panthera onca*. *Oryx*. 2018;52(2):300–15.
- Haag T, Santos AS, Sana DA, Morato RG, Cullen L Jr, Crawshaw PG Jr, et al. The effect of habitat fragmentation on the genetic structure of a top predator: loss of diversity and high differentiation among remnant populations of Atlantic Forest jaguars (*Panthera onca*). *Mol Ecol*. 2010 Nov; 19(22):4906–21.
- Colchero F, Conde DA, Manterola C, Chávez C, Rivera A, Ceballos G. Jaguars on the move: modeling movement to mitigate fragmentation from road expansion in the Mayan Forest. *Anim Conserv*. 2011;14(2):158–66.
- Polisar J, Brien TGO, Matthews SM, Beckmann JP, Sanderson EW, Rosas-Rosas OC, et al. Review of Jaguar Survey and Monitoring Techniques and Methodologies. 2014.
- Roques S, Sollman R, Jácomo A, Tôres N, Silveira L, Chávez C, et al. Effects of habitat deterioration on the population genetics and conservation of the jaguar. *Conserv Genet*. 2016;17(1):125–39.
- de la Torre JA, Núñez JM, Medellín RA. Spatial requirements of jaguars and pumas in southern Mexico. *Mamm Biol*. 2017;84:52–60.
- Paviolo A, De Angelo C, Ferraz KMPMB, Morato RG, Martinez Pardo J, Srebek-Araujo AC, et al. A biodiversity hotspot losing its top predator: the challenge of jaguar conservation in the Atlantic Forest of South America. *Sci Rep Nature Publishing Group*. 2016;6(37147):1–16.
- Romero-Muñoz A, Torres R, Noss AJ, Quiroga V, Thompson JJ, et al. Habitat loss and overhunting synergistically drive the extirpation of jaguars from the Gran Chaco. *Divers Distrib*. 2019;25(2):176–90.
- Roques S, Furtado M, Jácomo ATA, Silveira L, Sollmann R, Tôres NM, et al. Monitoring jaguar populations *Panthera onca* with non-invasive genetics: a pilot study in Brazilian ecosystems. *Oryx*. 2014;48(03):361–9.
- de la Torre JA, Núñez JM, Medellín RA. Habitat availability and connectivity for jaguars (*Panthera onca*) in the southern Mayan Forest: conservation priorities for a fragmented landscape. *Biol Conserv Elsevier Ltd*. 2017;206: 270–82.
- Eizirik E, Kim JH, Menotti-Raymond M, Crawshaw PG, O'Brien SJ, Johnson WE. Phylogeography, population history and conservation genetics of jaguars (*Panthera onca*, Mammalia, Felidae). *Mol Ecol*. 2001 Jan;10(1):65–79.
- Wulfsch C, Caragiulo A, Dias-Freedman I, Quigley H, Rabinowitz S, Amato G. Genetic diversity and population structure of Mesoamerican jaguars (*Panthera onca*): implications for conservation and management. *PLoS One*. 2016;11(10):e0162377.
- Ray DK, Welch RM, Lawton RO, Nair US. Dry season clouds and rainfall in northern Central America: implications for the Mesoamerican biological corridor. *Glob Planet Chang*. 2006;54(1–2):150–62.
- Rabinowitz AR, Nottingham JBG. Ecology and behaviour of the jaguar (*Panthera onca*) in Belize, Central America. *J Zool*. 1986;210:149–59.
- Figueroa OA. The ecology and conservation of jaguars (*Panthera onca* in Central Belize: conservation status, diet, movement patterns and habitat use (PhD thesis). University of Florida, Florida, USA; 2013.

17. Foster RJ. The ecology of jaguars (*Panthera onca*) in a human-influenced landscape (PhD thesis). University of Southampton, Southampton, UK; 2008.
18. Meerman J, Wilson JR, Andrade V, Taegar-panton T. The Belize National Protected Areas System Plan. Taskforce on Belize's Protected Areas Policy and System Plan. 2005. p. 26.
19. Harmsen BJ, Foster RJ, Silver S, Ostro LET, Doncaster. C. The ecology of jaguars in the Cockscomb Basin Wildlife Sanctuary, Belize. Biology and Conservation of Wild Felids. Oxford University Press; 2007. p. 403–16.
20. Kay E, Dickerson A, Urbina Y, Lizama D, Correa E, Cruz F, et al. Central Belize Corridor: Conservation Action Plan. 2015.
21. Petracca LS, Frair JL, Cohen JB, Calderón AP, Carazo-Salazar J, Castañeda F, et al. Robust inference on large-scale species habitat use with interview data: the status of jaguars outside protected areas in Central America. *J Appl Ecol*. 2018;55(2):723–34.
22. Mayer LR. Global conservation organizations applaud government of Belize for new commitment to protect Central America's largest, highly imperiled forest. Belize: Global Wildlife Conservation; 2019.
23. Rabinowitz A, Zeller KA. A range-wide model of landscape connectivity and conservation for the jaguar, *Panthera onca*. *Biol Conserv*. 2010;143(4):939–45.
24. Rouget M. Measuring conservation value at fine and broad scales: implications for a diverse and fragmented region, the Agulhas plain. *Biol Conserv*. 2003;112(1):217–32.
25. Wultsch C, Waits LP, Kelly MJ. Noninvasive individual and species identification of jaguars (*Panthera onca*), pumas (*Puma concolor*) and ocelots (*Leopardus pardalis*) in Belize, Central America using cross-species microsatellites and faecal DNA. *Mol Ecol Resour*. 2014;14:1171–82.
26. Wultsch C, Waits LP, Kelly MJ. A comparative analysis of genetic diversity and structure in jaguars (*Panthera Onca*), pumas (*Puma Concolor*), and ocelots (*Leopardus pardalis*) in fragmented landscapes of a critical Mesoamerican linkage zone. *PLoS One*. 2016;11(3):e0151043.
27. Foster RJ, Harmsen BJ, Doncaster CP. Habitat use by sympatric jaguars and pumas across a gradient of human disturbance in Belize. *Biotropica*. 2010; 42(6):724–31.
28. Cavalcanti SMC, Gese EM. Spatial ecology and social interactions of jaguars (*Panthera onca*) in the southern Pantanal, Brazil. *J Mammal*. 2009;90(4):935–45.
29. Harmsen BJ, Foster RJ, Sanchez E, Gutierrez-González CE, Silver SC, Ostro LET, et al. Long term monitoring of jaguars in the Cockscomb Basin wildlife sanctuary, Belize; implications for camera trap studies of carnivores. *PLoS One*. 2017;12(6):e0179505.
30. Palomares F, Roques S, Chávez C, Silveira L, Keller C, Sollmann R, et al. High proportion of male faeces in jaguar populations. *PLoS One*. 2012;7(12): e52923.
31. Antunes A, Troyer JL, Roelke ME, Pecon-Slattery J, Packer C, Winterbach C, et al. The evolutionary dynamics of the lion *Panthera leo* revealed by host and viral population genomics. *PLoS Genet*. 2008;4(11):e1000251.
32. Yumnam B, Jhala YV, Qureshi Q, Maldonado JE, Gopal R, Saini S, et al. Prioritizing tiger conservation through landscape genetics and habitat linkages. *PLoS One*. 2014;9(11):e111207.
33. Latch EK, Dharmarajan G, Glaubitz JC, Rhodes OE. Relative performance of Bayesian clustering software for inferring population substructure and individual assignment at low levels of population differentiation. *Conserv Genet*. 2006;7(2):295–302.
34. Manel S, Gaggiotti OE, Waples RS. Assignment methods: matching biological questions with appropriate techniques. *Trends Ecol Evol*. 2005; 20(3):136–42.
35. Evanno G, Regnaut S, Goudet J. Detecting the number of clusters of individuals using the software STRUCTURE: a simulation study. *Mol Ecol*. 2005;14(1):2611–20.
36. Chen C, Durand E, Forbes F, François O. Bayesian clustering algorithms ascertaining spatial population structure: a new computer program and a comparison study. *Mol Ecol Notes*. 2007;7(5):747–56.
37. Lonsinger RC, Schweizer RM, Pollinger JP, Wayne RK, Roemer GW. Fine-scale genetic structure of the ringtail (*Bassariscus astutus*) in a Sky Island mountain range. *J Mammal*. 2015;96(2):257–68.
38. Chavez C, Ceballos G, Medellín R, Zarza H. Primer Censo Nacional del Jaguar. Conservación y manejo del jaguar en México: estudios de caso y perspectivas. Mexico City: CONABIO-UNAM-ALIANZA WWF TELCEL; 2007. p. 133–42.
39. Conde D a, Colchero F, Zarza H, Christensen NL, Sexton JO, Manterola C, et al. Sex matters: Modeling male and female habitat differences for jaguar conservation. *Biol Conserv Elsevier Ltd*; 2010;143(1):1980–1988.
40. Thornton D, Zeller K, Rondinini C, Boitani L, Crooks K, Burdett C, et al. Assessing the umbrella value of a range-wide conservation network for jaguars (*Panthera onca*). *Ecol Appl*. 2016;26(4):1112–24.
41. Cullen L, Stanton JC, Lima F, Uezu A, Perilli MLL, Resit AH. Implications of fine-grained habitat fragmentation and road mortality for jaguar conservation in the Atlantic forest, Brazil. *PLoS One*. 2016;11(12):e0167372.
42. Srbek-Araujo AC, Haag T, Chiarello AG, Salzano FM, Eizirik E. Worrisome isolation: noninvasive genetic analyses shed light on the critical status of a remnant jaguar population. *J Mammal*. 2018;99(2):397–407.
43. Espinosa S, Celis G, Branch LC. When roads appear jaguars decline: increased access to an Amazonian wilderness area reduces potential for jaguar conservation. *PLoS One*. 2018;13(1):e0189740.
44. Milanese P, Holderegger R, Caniglia R, Fabbri E, Galaverni M, Randi E. Expert-based versus habitat-suitability models to develop resistance surfaces in landscape genetics. *Oecologia*. 2017;183(1):67–79.
45. Balkenhol N, Holbrook JD, Onorato D, Zager P, White C, Waits LP. A multi-method approach for analyzing hierarchical genetic structures: a case study with cougars *Puma concolor*. *Ecography (Cop)*. 2014;37(6):552–63.
46. Zeller KA, McGarigal K, Whiteley AR. Estimating landscape resistance to movement: a review. *Landsc Ecol*. 2012;27(6):777–97.
47. Milanese P, Holderegger R, Bollmann K, Guggerli F, Zellweger F. Three-dimensional habitat structure and landscape genetics: a step forward in estimating functional connectivity. *Ecology*. 2017;98(2):393–402.
48. Allendorf FW, Hohenlohe PA, Luikart G. Genomics and the future of conservation genetics. *Nat Rev Genet*. 2010;11(10):697–709.
49. Kocher TD, Thomas WK, Meyer A, Edwards SV, Pääbo S, Villablanca FX, et al. Dynamics of mitochondrial DNA evolution in animals: amplification and sequencing with conserved primers. *Proc Natl Acad Sci*. 1989;86(16):6196–200.
50. Farrell LE, Roman J, Sunquist ME. Dietary separation of sympatric carnivores identified by molecular analysis of scats. *Mol Ecol*. 2000;9(10):1583–90.
51. Kitano T, Umetsu K, Tian W, Osawa M. Two universal primer sets for species identification among vertebrates. *Int J Legal Med*. 2007;121(5):423–7.
52. Jukes TH, Cantor CR. Evolution of protein molecules. *Mamm Protein Metab*. 1969;33(18):21–123.
53. Menotti-Raymond M, David VA, Lyons LA, Schäffer AA, Tomlin JF, Hutton MK, et al. A genetic linkage map of microsatellites in the domestic cat (*Felis catus*). *Genomics*. 1999;57:9–23.
54. Covarrubias-Pazarán G, Díaz-García L, Schlautman B, Salazar W, Zalapa J. Fragman: an R package for fragment analysis. *BMC Genet*. 2016;17:62.
55. Galpern P, Manseau M, Hettinga P, Smith K, Wilson P. Allelmatch: an R package for identifying unique multilocus genotypes where genotyping error and missing data may be present. *Mol Ecol Resour*. 2012;12(4):771–8.
56. Waits LP, Luikart G, Taberlet P. Estimating the probability of identity among genotypes in natural populations: cautions and guidelines. *Mol Ecol*. 2001; 10(1):249–56.
57. Van Oosterhout C, Hutchinson WF, Wills DPM, Shipley P. MICRO-CHECKER: software for identifying and correcting genotyping errors in microsatellite data. *Mol Ecol Notes*. 2004;4:535–8.
58. Jombart T. ADEGENET: a R package for the multivariate analysis of genetic markers. *Bioinformatics*. 2008;24:1403–5.
59. Goudet J. HIERFSTAT, a package for R to compute and test hierarchical F-statistics. *Mol Ecol Notes*. 2005;5(1):184–6.
60. Rousset F. GENEPOP'007: a complete re-implementation of the GENEPOP software for windows and Linux. *Mol Ecol Resour*. 2008;8(1):103–6.
61. Peakall R, Smouse PE. GENALEX 6: genetic analysis in excel. Population genetic software for teaching and research. *Mol Ecol Notes*. 2006;6(1): 288–95.
62. Pritchard JK, Stephens M, Donnelly P. Inference of population structure using multilocus genotype data. *Genetics*. 2000;155(2):945–59.
63. Francis RM. POPHELPER: an R package and web app to analyse and visualise population structure. *Mol Ecol Resour*. 2017;1:27–32.
64. Durand E, Jay F, Gaggiotti OE, François O. Spatial inference of admixture proportions and secondary contact zones. *Mol Biol Evol*. 2009;26(9):1963–73.
65. Guillot G, Mortier F, Estoup A. GENELAND: a computer package for landscape genetics. *Mol Ecol Notes*. 2005;5(3):712–5.
66. Lee C, Abdoool A, Huang C-H. PCA-based population structure inference with generic clustering algorithms. *BMC Bioinformatics*. 2009;10(Suppl 1):S73.
67. Jombart T, Devillard S, Balloux F, Faloux D, Stephens M, Pritchard J, et al. Discriminant analysis of principal components: a new method for the analysis of genetically structured populations. *BMC Genet*. 2010;11(1):94.

68. Pew J, Muir PH, Wang J, Frasier TR. RELATED: an R package for analysing pairwise relatedness from codominant molecular markers. *Mol Ecol Resour.* 2015;15(3):557–61.
69. Shah VB, McRae BH. Circuitscape : A Tool for Landscape Ecology. Proceedings of the 7th Python in Science Conference. 2008. p. 62–5.
70. Meerman J, Clabaugh J. Biodiversity and environmental resource data system of Belize (BERDS) [internet]. Online. 2016; Available from: <http://www.biodiversity.bz>.
71. Jędrzejewski W, Robinson HS, Abarca M, Zeller KA, Velasquez G, Paemelaere EAD, et al. Estimating large carnivore populations at global scale based on spatial predictions of density and distribution - application to the jaguar (*Panthera onca*). *PLoS One.* 2018;13(3):1–25.
72. Wang J. Triadic IBD coefficients and applications to estimating pairwise relatedness. *Genet Res.* 2007;89(3):135–53.
73. Astete S, Sollmann R, Silveira L. Comparative ecology of jaguars in Brazil. *Cat News Spec Issue.* 2007;4:9–14.
74. Zarza H, Chavez C, Ceballos G. Uso de habitat del jaguar a escala regional en en paisaje dominado por actividades humanas en el sur de la península de Yucatan. *Conservación y manejo del jaguar en México: estudios de caso y perspectivas.* CONABIO –U. Mexico City; 2007. p. 101–110.
75. Zarco-González M, Rodríguez-Soto C, Monroy-Vilchis O, Urios V. Cougar and jaguar habitat use and activity patterns in Central Mexico. *Anim Biol.* 2009; 59(2):145–57.
76. de la Torre JA, Medellín RA. Jaguars *Panthera onca* in the greater Lacandona ecosystem, Chiapas, Mexico: population estimates and future prospects. *Oryx.* 2011;45(4):546–53.
77. Chávez C, Zarza YH. Distribución potencial del habitat del jaguar y areas de conflicto humano-jaguar en la península de Yucatan. *Rev Mex Mastozool.* 2009;13(1):46–62.
78. Figel JJ, Botero-Cañola S, Forero-Medina G, Sánchez-Londoño JD, Valenzuela L, Noss RF. Wetlands are keystone habitats for jaguars in an intercontinental biodiversity hotspot. *PLoS One.* 2019;14(9):e0221705.
79. Santos F, Carbone C, Wearn OR, Rowcliffe JM, Espinosa S, Moreira MG, et al. Prey availability and temporal partitioning modulate felid coexistence in Neotropical forests. *PLoS One.* 2019;14(3):e0213671.
80. Queller DC, Goodnight KF. Estimating relatedness using genetic markers. *Evolution (N Y).* 1989;43(2):258–75.
81. Li CC, Weeks DE, Chakravarti A. Similarity of DNA fingerprints due to chance and relatedness. *Hum Hered.* 1993;43(1):45–52.
82. Ritland K. Estimators for pairwise relatedness and individual inbreeding coefficients. *Genet Res.* 1996;67(02):175–85.
83. Lynch M, Ritland K. Estimation of pairwise relatedness with molecular markers. *Genetics.* 1999;152(4):1753–66.
84. Wang J. An estimator for pairwise relatedness using molecular markers. *Genetics.* 2002;160(3):1203–15.
85. Milligan BG. Maximum-likelihood estimation of relatedness. *Genetics.* 2003; 163(3):1153–67.
86. Stori FT, Peres CM, Turra A, Pressey RL. Traditional ecological knowledge supports ecosystem-based Management in Disturbed Coastal Marine Social-Ecological Systems. *Front Mar Sci.* 2019;6(September):1–22.
87. Assis JC, Giacomini HC, Ribeiro MC. Road permeability index: evaluating the heterogeneous permeability of roads for wildlife crossing. *Ecol Indic Elsevier.* 2019;99(1):365–74.
88. Stricker HK, Gehring TM, Donner D, Petroelje T. Multi-scale habitat selection model assessing potential gray wolf den habitat and dispersal corridors in Michigan, USA. *Ecol Modell Elsevier.* 2019;397(1):84–94.
89. Drescher M, Perera AH, Johnson CJ, Buse LJ, Drew CA, Burgman MA. Toward rigorous use of expert knowledge in ecological research. *Ecosphere.* 2013;4(7):1–26.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

